UNIVERSITÀ POLITECNICA DELLE MARCHE

DIPARTIMENTO SCIENZE DELLA VITA E DELL'AMBIENTE

Laurea magistrale in

Biologia Marina

Biodiversità e driver ambientali delle comunità ittiche nei differenti habitat delle coste Calabresi

Biodiversity and environmental drivers of fish assemblages in different habitats of the Calabrian coasts

Tesi di Laurea di: di:

Enrico Cecapolli

Docente Referente Chiar.mo Prof.

Emanuela Fanelli

Correlatore

Antonio Di Franco

Sessione Straordinaria (febbraio 2023)

Anno Accademico 2021/2022

ABSTRACT

La stima della biodiversità è uno degli aspetti chiave per descrivere e preservare la diversità e funzionamento degli ecosistemi. La biodiversità degli ecosistemi marini e la distribuzione delle specie è modellata da una moltitudine di fattori che interagiscono tra loro, e negli ultimi decenni è stata osservata una riduzione della diversità specifica, con conseguenze su interi ecosistemi e sulle loro interazioni con gli esseri umani. In questo studio è stata applicata una metodologia di visual census per quantificare la diversità specifica della fauna ittica delle coste della Calabria, andando ad approfondire diverse sue sfaccettature, focalizzandosi sui popolamenti specifici degli habitat costieri (habitat rocciosi, sabbiosi e praterie di *Posidonia oceanica*). In particolare, sono state analizzate proprietà come la stima delle specie presenti, le loro interazioni e le differenze nelle comunità tra i vari habitat. Da queste osservazioni è emersa una maggior ricchezza specifica in habitat rocciosi, ma un contributo fondamentale di tutti gli habitat, quantificato con la capacità di portare un alto contributo alla β-diversità, specialmente da parte delle comunità sabbiose, nonostante la loro minor ricchezza specifica rispetto agli altri habitat. Anche le specie che contribuiscono maggiormente a creare queste differenze tra gli habitat sono state analizzate, con risultati che rispecchiano un alto contributo delle specie abitanti gli habitat sabbiosi, ma anche diffusa presenza di specie generaliste e presenti in habitat rocciosi e/o praterie di *P. oceanica*. Inoltre, è stata modellata una predizione della ricchezza specifica su tutta l'estensione delle coste della Calabria, basata sui dati ottenuti durante i campionamenti, ottenendo una ricchezza specifica generalmente bassa su tutta la fascia costiera, ma caratterizzata da alcuni hotspot di diversità in aree che presentano elevata eterogeneità riguardante gli habitat.

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5. REFERENCES

1. INTRODUCTION

1.1. The concept of biodiversity

Biodiversity, defined as the "variability among living organisms from all sources, including, inter alia, terrestrial, marine, and other aquatic ecosystems, and the ecological complexes of which they are part: this includes diversity within species, between species and of ecosystems" (UN Convention on Biological Diversity, 1992), has been highlighted in the last few decades as one of the key features to describe and understand the nature and functioning of ecosystems (Loreau et al., 2001; Midgley, 2012). The continuous challenges that natural systems are facing worldwide is requiring a deeper understanding of the attributes characterizing biodiversity, in order to better assess and manage the ecological consequences of changes occurring within those systems, also in relation to the Nature's Contribution to People (NCP, Díaz et al., 2018), a recent concept that expand the one of Ecosystem Services (ES) provided by nature to humans (Bagousse-Pinguet et al., 2019). The NCP refers to "all the benefits and detriments that people get from their relationships with the rest of the living world" (Hill et al., 2021). In fact, while the ES are the way by which people benefit from what the natural environment is able to provide them directly or indirectly and can be affected by every change occurring in the ecosystems themselves, deeply affecting the well-being of people all over the

world (Worm et al., 2006), the concept of NCP is wider and less economically driven, including many aspects which can have both positive and negative effects on humans. In broad terms, the NCP can be divided in three categories, material, non-material, and regulating NCP (Hill et al., 2021).

There are a multitude of processes shaping species diversity and distribution in marine systems. Among the best known and acknowledged ones there are external drivers acting on them, both biotic (e.g. competition, predation; Van der Putten et al., 2010) and abiotic (e.g. climate and oceanographic features; Pearson et al. 2003). These drivers can act together in a synergistic way (Benton, 2009; Lewis et al., 2017), to model the structure and functions of marine ecosystems. There is still an ongoing debate on how such drivers contribute and the spatial scale at which they are more relevant in influencing biodiversity (Araújo and Luoto, 2007; Meier et al., 2010; Wiens, 2011; Lewis et al., 2017), but their overall importance is widely recognized. The general trend of the last decades has been a reduction in species diversity (McCauley et al., 2015) and in some cases even the loss of entire functional groups, hampering the ability of ecosystems to deliver ES to human communities, especially in some complex, delicate, and poorly known marine environments (e.g. the deep sea) (Danovaro et al., 2008).

1.2. Fish as biodiversity indicators

In marine ecosystems, fishes have received a special focus as they are often considered as a proxy of ecosystem biodiversity, because they represent a highly diverse group, cover a wide range of ecological functions, and their taxonomy and biological and ecological traits are relatively well known (Rice, 2003; Stuart-Smith et al., 2013; Duffy et al., 2016). Fish are among the most common organisms in the world's oceans, and overall the most diverse group of vertebrates, inhabiting all the possible aquatic ecosystems (Manel et al., 2020). Fish can exhibit a multitude of shapes and growth forms, from elongated and adapted for fast swimming, to flattened and suitable to camouflage under soft bottoms, but including an uncountable number of other forms standing in the middle. Such organisms optimized their biology over millions of years of evolution, acquiring the capability to survive in almost every ecosystem. Fish species are able to express a variety of life habits, differentiating from each other for reproductive, feeding, and behavioral features, depending on the conditions they live in and their life history.



Figure 1.1. (From Manel et al 2020). Genetic diversity of marine fish in the world, estimated as the mean number of mutations per base pair for Cytochrome Oxidase Subunit 1 sequence across species (a) 514 cells for marine fishes. (b) Genetic diversity averaged across cells within latitudinal band of 10° and is plotted as a function of latitude for marine fish species.

The general classification under the name "fish" includes both cartilaginous fishes, named **Chondrichthyes** (i.e. sharks, rays, etc.), and bony fishes, named **Osteichthyes**, representing the vast majority of the total number of species (Frasca et al., 2018). Osteichthyes can be further divided into ray-finned organisms, or Actinopterygii, and lobe-finned fish, known as Sarcopterygii. The first bony fish currently identified has dated back around 425 million years (Zhao et al., 2021), and this can give us an idea of the longevity of this clade and the amount of evolutionary history they have been through, resulting in the species present now in our oceans with all the adaptations that they possibly exhibit to survive in very disparate conditions.

Many aspects of fish biology and biodiversity are still unknown, especially in extreme or remote habitats, but many times even the most accessible

environments can be associated with knowledge gaps. Recent estimates are of around 15.000 to 20.000 fish species for marine ecosystems, unevenly distributed and mainly concentrated in the shallow coastal ecosystems (within the first 200 m of depth) (Henseler et al., 2019). This is firstly due to more favorable living conditions, giving the presence of **light** (and so the presence of primary producers at the bottom of the trophic webs, that sustain all the higher levels), levels of **hydrostatic pressure** compatible with life (which could be instead a limiting factor in deep-sea ecosystems; Yancey et al., 2014), along with many other factors. A considerable part is also related to an evident gap in our knowledge regarding the deeper portions of the world's oceans, which needs to be addressed in future investigations (Ramirez-Llodra et al., 2011).

Organisms belonging to this taxon can therefore represent a very useful proxy to assess the biodiversity of marine environments effectively and rapidly, thanks to the relatively high knowledge of their diversity and ecological characteristics compared to other less known groups.

1.3. Assessing biodiversity

For the reasons above-described, it appears necessary to move forward the knowledge about biodiversity in order to properly assess the diversity of an

area, but also to highlight multiple shades of it, completing the evaluation by using multiple aspects and estimates, including the **species richness**, meaning the number of species in a given area (Solan et al., 2012; Gotelli et al., 2013), and species abundance, which is instead the number of fish individuals for a species in a given area (Verberk, 2011), giving the possibility to assess diversity using a compound approach, that provides more accurate answers to our questions (Stuart-Smith et al., 2013). For example, it is widely known that biodiversity generally positively influences the productivity and the stability of a system, enhancing the possible usage spectrum of the available resources (Cardinale et al., 2011). But different ecosystems react in different ways and with different magnitude to the oscillation in biodiversity that can be driven both by natural and anthropogenic stressors (Hooper et al., 2005), highlighting the necessity to increase knowledge on such relevant issues, to finally enhance the power to make inference regarding this complex topic. Furthermore, the consideration of more than one aspect is a good way to reduce biases, which is a critical issue to account for when we aim to obtain biodiversity estimates. Moreover, to assess the diversity of an area under investigation, as in any kind of scientific approach to the test of a hypothesis, there is a need to perform replicates of the measurement of interest (e.g. diversity). Replicates are repeated measurements of the same sample that represent independent

measures of the random noise associated with protocols or equipment (Blainey et al., 2014). Replication provides a gain of information, which justifies the costs related to the increased effort for the measurements (Dennis et al., 2010). More specifically, among the possible measurements related to biodiversity, also a large set of diversity indices focusing on the **taxonomical** dimension of biodiversity is available. A **diversity index** is a quantitative measure that reflects how many species (or other biological entities) there are in a given community and that can consider different aspects of the relations between the considered species.

Among the possible indices, one of the most used in marine ecology is the **Shannon-Wiener index** (Loiseau et al., 2015), which allows to assess the evenness of species in a community, while emphasizing the diversity component of that community (Shannon and Weaver, 1949). Another widespread used metric is the **Simpson index**. This index is a dominance index giving more consideration to common or dominant species of a community (Simpson, 1949).

Beside considering the diversity of a specific area, which is also known as α diversity (Magurran, 2004), a complete and comparative spatial evaluation should also include β -diversity, which refers to a measure of the difference in species composition either between two or more local assemblages or between

local and regional assemblages (Koleff et al., 2003). Using such scale of the species diversity allows to efficiently compare the communities inhabiting the different habitats or sampling units. The β -diversity can be also considered as driven by different components to highlight the differences between the sites, which are both regarding the spatial differences between each replicate. This measure can inform on the "uniqueness" of each individual site relative to the entire seascape community, also known as Local Contribution to Beta Diversity (Legendre & de Cáceres, 2013), and the Species contribution to Beta Diversity (Legendre & de Cáceres, 2013), which is a different way of considering the differences in diversity based on the species rather than on spatial entities such as sites, still relevant for contributing to the heterogeneity of the seascape (Harper et al., 2022). This kind of approach could be a powerful instrument that allows to highlight interesting patterns in a comparative spatial investigation.

Also, other very useful descriptors that can be assessed are the species in common among all the different areas or habitats, or just between some of them, and the species present in just one habitat ("unique species"), which are good quantitative estimators as well of the degree of change between them, giving a more detailed picture about the shades and the gradients that biological diversity is formed by (Harper et al., 2022).

To enhance the potential to estimate the biodiversity of the community under study, a sound assessment should also include **biomass** measurements, defined as the total wet weight of individuals by area, that could be a possible descriptor to estimate the actual stability of a community (Cardinale et al., 2013). Regarding biomass, marine ecology assessments usually elaborate information to obtain a group's or population's biomass, which are historically of interest to ecologists and resource managers (Duffy et al., 2016). Biomass increases the capacity to evaluate the actual quantity of organisms, as it also considers the size of the organisms rather than just their presence as individuals (Fiorella et al., 2019). Furthermore, the consideration of the physical parameters in its calculation (e.g. length), biomass is also a useful metric for quantifying taxa and/or species whose members are of very different sizes (Bar-On et al., 2018), and so to give a very "readable" parameter for the assessment of the actual fish presence in an area or habitat.

The macroscopic nature of fish allows a non-destructive approach in their assessment, enabling the possible application of methods associated with reduced disturbance to species and ecosystems. This could be important when thinking about the eventual use of such organisms in the evaluation areas under any restriction or of particular biological interest, or just in order to improve the sustainability of this kind of studies. The best techniques to put these

theoretical concepts into practical terms are those which rely on visual tools, such as **Underwater Visual Census** (UVC), **Baited Underwater Videos** (BUVs), and related procedures (Colton et al., 2010; Caldwell et al., 2016). These techniques can be complementary for several reasons, such as the different operative depths at which they can be applied, and the species targeted by each method (Lowry et al., 2012).

In fact, literature suggests that UVC holds the potential to sample a wide spectrum of species, including more cryptic ones, while BUVs sampling



Figure 1.2. Two fishes sampled over the study. Top-left: Scorpaena porcus; bottom-right: Apogon imberbis

usually majorly focuses on the predatory fishes (Lowry et al., 2012; Cheal et

al., 2021), due to the presence of attractive bait.

These types of data should ideally be combined to fishery-dependent data, that can give information on wider spatial and temporal scales but are generally less precise regarding the location of the fishing ground and the classification of species caught (Pennino et al., 2016), while fishery-independent data (or survey-based methods, e.g. UVC) have a more limited spatial and temporal extension, leading to a lower quantity of data but potentially higher quality, because of the specific sampling design of the study (Hilborn et al., 2013). The synergistic use of the two approaches allows to obtain more continuous, realistic, and robust assessments over time (Aglieri et al., 2021), but the use of survey-based methods (e.g. UVC and BUV) already effectively combines both quantitative and qualitative results if properly performed and appears to be a useful approach to start estimating poorly known communities.

The habitat-based characterization of fish diversity can possibly have a good potential to assess the biodiversity of an area, because the differences found in biodiversity (both α - and β -diversity) for each habitat can inform on the importance of underrated portions of complex ecosystems, composed by more than just one habitat. Furthermore, the connectivity between the habitats is another aspect to account for when trying to achieve sound protection of species, especially if quite highly mobile organisms with a complex ecology such as fishes are considered. It is known that different life stages of fishes can

occur in different habitats: for example, seagrass meadows are well known to be nursery areas (Madi Moussa et al., 2020), hosting a number of juveniles that will then move to other habitats when they become adults, because they offer a very good shelter, and stable and productive environment for them to increase their survival chances during the most delicate period of their life. This suggests that habitats should not be considered as isolated compartments, but more on their completeness, to have a more comprehensive estimate of their true contribution to the overall diversity.

1.4. Aim of the study

The overall aim of this study is to assess pattern of coastal fish diversity and its drivers in different habitats. To do so, we used as a case study the coastal waters of the Calabria Region, Southern Italy (see a description in Methods section 2.1.).

The investigation proceeded following a multiple and progressive stepped approach, developed around three main goals:

- 1. Evaluate the fish diversity of each habitat under consideration.
- 2. Assess the **contribution of each habitat** on the overall α -diversity.
- 3. Scale up the biodiversity assessment, especially based on how the biotic and abiotic drivers can shape fish assemblages.

More specifically, the first goal is to evaluate the fish diversity of each habitat under consideration (i.e. rocky, sandy, Posidonia meadows, see Section 2.1 for further details), possibly providing useful information for future assessments of similar habitats and enhance the comparative potential with already present investigations, even using different approaches about the habitat-based fish diversity assessments (Thiriet et al., 2016; Clarkson et. al., 2021; Arndt et al., 2018; Bolgan et al., 2022). It is believed indeed, also based on previous studies (Guidetti, 2000; Gratwicke et al., 2005), that each habitat hosts a specific diversity, composed of many exclusive species that rely on food sources present or that developed the ability to use resources of the habitat where they live, also depending on the biotic and abiotic drivers acting on the habitat, and so all of them deserve adequate consideration. This does not only mean the simple assessment of species richness, which remains still very important but also to check which species are present and how their niches could possibly overlap and how could they interact, to see the specific potential of every area under study.

The second expected goal is to "use" the obtained knowledge about the habitatspecific fish biodiversity to assess the contribution of each habitat on the overall α -diversity. In fact, this is still one of the first facets of biodiversity analyzed on a small scale investigation, and a key feature of an environment to then

expand to other components of diversity (e.g. β -diversity; Lazzari et al., 2020). The relative contribution of a habitat to the α -diversity is also a good measure of the potential of every habitat to provide diversity to a larger system and its specific contribution to people (Díaz et al., 2018), especially in coastal habitats (Henseler et al., 2019), being the first step to investigate the connectivity between the habitats and their relative value in determining the overall biodiversity of a larger area (e.g. the Calabrian coasts), that can be expressed as γ -diversity (Whittaker, 1960).

Regarding this, scaling up the biodiversity assessment on the specific study locations, based on how the drivers can shape fish assemblages and the possible changes occurring in the ecological relationships between them becomes fundamental.

The final aim of this process is to increase the knowledge not only regarding the habitats, but also concerning the potential of this important portion of the Mediterranean Sea in contributing to the spatial distribution of biodiversity in the entire basin (Edgar et al., 2016).



Figure 1.3. An UVC (Underwater Visual Census) transect performed on a sandy habitat (source: EC).

The last step will be subsequently the prediction of the distribution of the fish species with a modeling approach, that allows to expand the spatial scale in which the species presence can be assessed. In fact, this could inform on any possible hotspot of biodiversity, and eventually on other areas with lower values of fish diversity, (i.e. "coldspots"), but also areas with rare or vulnerable species that should be managed, especially if a reduction in their density is noticed.

To do so, both data taken from the field (e.g. depth) and derived from online

databases (e.g. temperature, salinity) are of vital importance to correlate possible fish diversity patterns found to biotic and abiotic drivers (see Section 1.1 for more detailed information) in the different habitats. Their importance is relevant also considering the different spatial units, to model fish distribution in the whole Calabrian coastline and try to develop a predictive distribution model (Rodrigues et al., 2022) of the considered fish species.

The findings of this study could inform management and conservation strategies, such as the 30x30 commitment that will likely engage countries in protecting 30% of their waters (with at least 10% of strictly protected waters, European commission, 2020). In fact, it is widely recognized that protected areas are one of the foundations for the conservation of biodiversity (Bleeker et al., 2011). However, at present time, the portion of adequately protected sea surface is insufficient and not evenly distributed (Claudet et al., 2020), so immediate efforts are needed to reach the aimed goals.

More specifically, identifying patterns of diversity and locating diversity hotspots could help guiding site selection for the implementation of new Marine Protected Areas (MPAs) and designing sound management strategies. This means that, if necessary, reduced fishing pressure, as well as other management initiatives, could be suggested to prevent the occurrence of possibly irreversible effects both on the ecosystem and in the services provided to people.

2. MATERIAL and METHODS

2.1. Study area

2.1.1. The Mediterranean Sea and the basins surrounding the Calabrian coastlines

The Mediterranean Sea covers an area of slightly over 2.5 million km² and it is a semi-enclosed basin, with the only openings to connect with the other oceans being represented by the Gibraltar Strait on the western side, where Atlantic water flows (Poulos, 2020), and the Suez Canal on the eastern side, opened in 1869 and connecting the Mediterranean to the Red Sea. The Suez Canal was opened to facilitate the commercial routes from Asia, but created a number of different new threats for the native biodiversity, such as the increased extinction rate (Gurevitch et al., 2004; Bellard et al., 2016) resulting from this connection (Zenetos et al., 2017). Average depth of Mediterranean Sea is ~1547 m (Barale, 2008), which is quite low compared to other world's oceans, averaging \sim 3850 m. Despite of its relatively small extension, the Mediterranean Sea hosts a large number of inhabitants surrounding it, divided into 28 different countries that belong to three continents (Europe, Africa, and Asia). Thus, also the anthropogenic impact level on the marine environment is high compared to larger basins around the world. Nonetheless, because of its intrinsic natural and

historical features, this basin is well known as a biodiversity hotspot (Coll et al., 2010), and in particular, the Calabrian coasts present of a variety of habitats and bio-oceanographic conditions. The biodiversity potential of this area is thus really high, but robust assessments need to be made to characterize fish diversity, including comprehensive data about the fish assemblages of the area. In fact, partial, fragmented, and often just catch-based data are present about this area (Busalacchi, 2010; Carlucci et al., 2018), not covering all the coastline extension (Tyrrhenian and Ionian Seas, Messina Strait).

This thesis was based on a large-scale survey which encompassed the Calabrian coasts, Southern Italy, central Mediterranean area. The Calabria Region has a marked peninsular shape and coastlines extend for almost 800 kilometers. In particular, the sampling design included five main locations, where the team could take fast boats from to reach the points chosen for sampling activities, selected also to reduce sampling constrains due to logistic issues.

The locations under study represent very important and strategic sites as they are at the center of the biogeographic border between the Eastern and the Western Mediterranean (Nicolaidou et al., 2012; Sperone et al., 2012). Overall, the three basins surrounding Calabria are: The Western Ionian Sea, the Messina Strait, and the Southern Tyrrhenian Sea.

The Ionian Sea is an extremely important region in terms of biodiversity and

coastal geomorphological heterogeneity (Galli et al., 2022), located between the Western basin limit and the southernmost portion of the Adriatic region, shaping its unique geomorphological features, such as some of the deepest trenches of the entire Mediterranean and a marked interannual variability in its conditions (Kalimeris et al., 2020). This reflects the high diversity potential of this sector resulting from variable abiotic and biotic drivers that can change their relative contribution over time (Benton et al., 2009; Meier et al., 2010). Also, negative aspects must be considered when discussing such various influxes, for example, the high number of Non-Indigenous Species (NIS) recorded in this area, majorly coming from the eastern basin through the Suez Canal, and also known as Lessepsian species. Some of these species are currently jeopardizing native fish assemblages and entire ecosystem dynamics all over the Mediterranean Sea (Carlucci et al., 2018), so assessing and knowing how widespread the problem is in this area could be crucial to design sound management strategies.

The **Messina Strait** is probably one of the most peculiar spots of the entire Mediterranean basin, connecting the Tyrrhenian and the Ionian Sea while separating Sicily from the Italian mainland. The area is characterized, as a consequence of the hydrological properties, by upwelling phenomena of the Levantine Intermediate Waters (LIW), coming straight from the Eastern Mediterranean and passing through the Ionian Sea, and finally resulting in a large amount of organic matter in addition to that arriving from the shallower layers. This leads to strong current flows and a steep slope of the sea bottom, which are factors that widely affect the distribution and presence of fish assemblages, both larval and adult stages (Rattray et al., 2016). The biological result of the abiotic factors acting is a high biodiversity and abundance of species, in many cases very similar to the Atlantic fauna, due to the resemblance of the oceanographic conditions (Spanò et al., 2017).

The last sector on focus is the **Tyrrhenian** portion of the Calabrian coasts, also characterized by high oceanographical and environmental complexity along the coastlines considered (Iacono et al., 2021). Due to these features, it supposedly holds some very interesting features that shape the regional diversity of its faunal assemblages, like the occurrence of many seamounts and the persistent influence of the Messina Strait currents (Busalacchi et al., 2010).

2.1.2. The sites

The sampling sites have been chosen from a wider set of potential sites, in order to cover the maximum geographic range possible in relation to the environmental gradients present in the area and increase the range of values associated to the predictors that could shape biodiversity patterns along the Calabrian coasts. The original set of potential sites was compiled in QGIS software (version 3.24.2) using data from the EUNIS (European Nature Information System) database. These data provide information on European habitats and has been a key tool to have reliable information on a guite wide spatial scale. The cells have been designed using a grid with a resolution of ~ 4 km, correspondent to the spatial division of the Copernicus Marine Environment Monitoring Service (CMEMS, http://marine.copernicus.eu) environmental data, that allowed to maximize the spatial range covered in relation to the environmental data correspondent to each cell. At the end of this process, each site was associated to a habitat-type. Nonetheless, before starting the sampling process, an echo sonar was used to precisely assess the depth and the kind of bottom, as well as a surficial visual inspection by snorkel, also known as ground-truthing. Any discrepancy from the original habitat of the sampling design has been recorded and replaced from the previous one, to be used in all the following analyses performed in the study.

2.2. The habitats of the Calabrian coasts

The extreme variety in the biotic and abiotic characteristics of the Calabrian coastal waters results in three possible categories of habitats that can be found (at the depths < 25 m). These habitats can be listed as: rocky bottoms, seagrass meadows (mainly *Posidonia oceanica*), and sandy bottoms.

2.2.1. Rocky bottoms

Rocky bottoms are usually very heterogeneous and diverse, thanks to their complexity and the creation of micro-habitats that can be exploited by organisms, offering shelter to juveniles, and allowing the simultaneous presence of a wider spectrum of species (Guidetti, 2000), thus bringing to the maximization of biodiversity when dealing with fish assemblages.

Of course, the definition of rocky bottoms remains pretty general, because of the multitude of shapes and the degree of complexity that may fall under this category, as well as the various benthic assemblages that cover the abiotic portion of the seascape, so the possible variety of organisms that can be found



Figure 2.1. A typical Mediterranean rocky habitat (source: EC)

depends more on the specific conditions that change case by case (Di Franco et al., 2021).

2.2.2. Posidonia meadows

Posidonia oceanica is the most represented and renowned seagrass species of the Mediterranean basin. *Posidonia oceanica* is a plant, so it relies on light to grow, and it can be found within the photic zone of sandy and mixed sea bottoms (Marbà et al., 2014; Balestri et al., 2015).



Figure 2.2. A typical Mediterranean Posidonia habitat on rocky bottom (source: EC)

In some cases, *Posidonia* can have a patchy distribution with some of the patches embedded inside other habitats. The structures formed by its roots and rhizomes can stabilize the sediments, and this is one of its most important

characteristics, along with numerous other benefits resulting from the habitat it forms. Even if the species' decline has been widely reported over the last decades (Chefaoui et al., 2018) mainly due to anthropogenic pressures, including the spread of invasive species (Telesca et al., 2015), Posidonia still has an extremely important role in providing services and represent a suitable habitat for multiple life stages of many organisms, especially mollusks and fishes. In fact, it is particularly considered a sensitive habitats used as nursery areas by many species, including those of commercial value. Being a nursery area, this habitat has a high seasonal variability in abundance and diversity of the associated fauna, which needs to be accounted for when performing biodiversity studies. The high sensitivity of Posidonia to a number of abiotic and biotic stressors (Pergen-Martini et al., 2006; Waycott et al., 2009; Telesca et al., 2015) requires immediate actions to quantify their potential to provide services and protect the associated diversity, and this can be done also by assessing the organisms that exploit this ecosystem and the relations occurring in it.

2.2.3. Sandy bottoms

Sandy bottoms are expected to be the less diverse habitats among the ones described in terms of specific richness (Guidetti, 2000; Schramm et al., 2021), because of the reduced heterogeneity due to the absence of spatial subdivisions over large surfaces, that can form micro-habitats like in the other cases, resulting in apparent monotonous bare substrates. Despite this, they are preferential homes for many fish species (e.g. *Lithognathus mormyrus*, *Xyrichtys novacula*, *Bothus podas*, etc.), using them also as feeding grounds or just transit areas (Guidetti, 2000), and other taxa such as mollusks. Soft bottoms can be the target for destructive anthropogenic activities, such as dredging and bottom trawling, and the consequences of those practices on the associated target and non-target fauna are potentially considerable.



Figure 2.3. A typical sandy bottom of the Mediterranean Sea (source: EC)

The investigation of soft bottoms assemblages is quite challenging because of the patchy distribution of organisms and the relative difficulties in detecting them, so most of the available data come from fishery-dependent information-FDI (Boudouresque et al., 2004). However, the use of visual methods, which has been rarely applied to sandy bottoms in the past, could be the key to better observing and correctly discriminating how different organisms interact, also based on their specific life habits.

2.3. Sampling locations and sites

Sampling locations included in the study were five: **Cetraro**, **Vibo Valentia**, **Capo Rizzuto**, **Amendolara**, and **Scilla**. These locations were chosen in order to cover different areas along Calabrian coasts, and also for logistic reasons as they are close to harbors, thus facilitating the access to boat for sampling. The total number of sampled sites was **62**, of which:

- 4 rocky sites
- 23 sandy sites
- 15 *Posidonia* sites



Figure 2.4. Map of the sampled sites (the colors are correspondent to the starting locations)

In particular, we sampled 14 sites in Cetraro, 11 in Vibo Valentia, 21 in Capo Rizzuto, 2 in Amendolara, and 14 in Scilla. The reduced number of sites in Amendolara was due to the smaller area suitable for UVC sampling (established during the sampling design at around – 25 m), despite the large area covered by the bank, i.e. the off-shore rocky bank "Secca di Amendolara", also included because of its potentially high biological interest. Capo Rizzuto is also an area of special interest for Calabrian waters, being the only Marine Protected Area (hereafter MPA) in the region. Our study did not aim to assess MPA effectiveness, but the area has been chosen because of its good fit with the sampling design.

2.4. Sampling method

The method chosen for collecting data on fish assemblages was **Underwater Visual Census** (UVC). UVC is a widespread technique used for non-invasive sampling of community-level data in shallow, nearshore habitats (Sale, 1980), including many techniques used to quantify fish populations (Thresher and Gunn, 1986). This sampling method was proven a reliable one in shallow waters to assess fish diversity, thanks to its unique features, i.e. short data processing times, accurate estimates for diversity, abundance, and specific traits of fish assemblages. UVC is comparable with other visual techniques (e.g. BRUVs), even if their complementarity enhances to a higher level their overall performances (Cheal et al., 2021).

The divers involved in sampling operations were five (ADF, MDL, AC, SG, EC), trained to perform UVC transects in the most consistent way possible and to ensure a correct estimation of fish abundances and sizes (Lowry et al., 2012) despite of the possible biases known for this technique, such as fish mobility, meteorological and water conditions of the site and the mentioned differences between the observers' knowledge and the critical judgment of assemblages. The depth range at which transects were carried out was between 0 m and around -25 m, which has been recognized as the maximum depth to complete

all the observations planned for every sampling day and remain within the safe

limits regarding SCUBA diving activities.

Within each of the 62 sites, 8 replicates (from now on referred to as "transects") were performed. In every site, one or two divers performed the transects, and the number of transects carried out by each operator has been assessed in the pre-diving briefing, based on many variables (depth of the site, number of dives already made during the day, etc.). Every operator was provided with a pencil, a blank sheet (on a slate) to annotate the information, and a reel to measure the transect. The divers proceeded in opposite direction to avoid covering the same areas over the replicates. At each transect, sampling was carried out according to the UVC (Underwater Visual Census) protocols described by Harmelin-Vivien et al. (1985), which has been commonly applied in the whole Mediterranean basin in the last years (Di Franco et al., 2009; Sala et al., 2012; Di Lorenzo et al., 2020). In particular, each transect was sampled using a **strip transect** of 25 m × 5 m, with a total surface of 125 m².
Along each transect, a trained diver operator swam one way at a constant speed (covering each transect in approximately 6–8 min) using SCUBA equipment and quantified several aspects of fish encountered (see below).



Figure 2.5. diver finishing a UVC transect (source: EC).

The transect was physically identified by a polyethylene line rolled up in a reel and ending with a small weight to keep it steady, with a fixed length of 25 m, to avoid any bias related to distances miscalculation underwater. The line was laid as the diver moved forward during its replicate to avoid excessive disturbance on fish (Dickens et al., 2011; Emslie et al., 2018).

Because of the cryptic nature of some benthic species in the Mediterranean assemblages (e.g. Blenniidae, Gobiidae, Scorpaenidae) (Thiriet et al., 2016),

during the way back to the starting point of each transect, the operator usually looked for any additional fish occurrence under rocks, in crevices and in other hidden portions of the sea bottom, to ensure a complete census of all species. For every transect, the diver also noted any deviation from the supposed habitat (noting it with a percentage of the dominant habitat).

2.4.1. Fish metrics measurements

During each transect, the diver recorded all fish species, estimating **abundance** whenever there was more than one individual at the same time, and the size of all fishes encountered. Actual number of fish encountered was recorded up to 10 individuals, whereas larger groups were recorded using categories of abundance (i.e. 11-30, 31-50, 51-200, 201-500, >500 ind.; see Harmelin-Vivien et al., 1985). Fish size (total length, TL) was recorded within 2 cm size classes for most of the species, and within 5 cm size classes for large-sized species (maximum size > 50 cm) such as *Epinephelinae*, accounting for a minimum level of visual error by the operator (Bortone and Mille, 1999; Edgar et al., 2004; Mallet and Pelletier, 2014).

2.4.2. Additional measurements

The diver also noted for each transect several other information to have a complete assessment of useful metrics to use in the data analysis process. Measures included some general information, i.e. **date** of the observation,

diver's name, and information about the site, such as its **code**, **latitude** and **longitude**, fundamental to geo-locating the observation, **initial** and **final depth**, that were averaged to obtain a mean depth for each transect. Lastly, the actual **type of habitat** sampled (Rocky, Sandy, *Posidonia*) was recorded, after visual confirmation of the EUNIS prediction. Any additional note about the transects was written too, to have it stored and available in any required case.

2.4.3. Environmental variables

Apart from all the variables recorded *in situ*, many others were taken from online databases, made available by Copernicus Marine Environment Monitoring Service (CMEMS).

The retrieved data included:

- Sea Surface Temperature (SST)
- Sea Bottom Temperature (SBT)
- Sea Surface Salinity (SSS)
- Concentration of **dissolved molecular oxygen** in sea water (O₂)
- Sea water **pH** (pH)
- Concentration of **ammonium** in sea water (NH₄)
- Concentration of **nitrates** in sea water (NO₃)
- Concentration of **phosphates** in sea water (PO₄)
- Concentration of **silicates** in sea water (Si)

• Concentration of **chlorophyll-a** in sea water (Chl)

Moreover, also the concentration of phytoplankton (PHYC) and the concentration of **zooplankton** (ZOOC), both expressed as carbon in sea water, were retrieved. The **bathymetries** were obtained from the EUNIS database, that has a higher resolution compared to other sources available, which is fundamental for the purposes intended in this kind of investigation, to have precise data about the depth of the shallow layers (< 30 m, sampled by UVC). The bathymetric data were used to obtain the **slope** and the **roughness** of the sea bottom, that could be useful variables to relate with fish diversity metrics. Finally, another driver considered was the human impact score, always spatially referenced and available from Micheli et al. (2013), which compounds estimates of 22 anthropogenic drivers (e.g. trawling, coastal population density, hypoxia, etc.) in the Mediterranean and the Black Sea. Each one of the data with a temporal variability has been handled by averaging the monthly values of the previous year at each location.

2.5. Data Handling and Analyses

All the subsequent analyses were conducted using Rstudio 2022.07.2 unless otherwise stated.

The abundances and biomasses were put into matrices in Microsoft Excel

(Version 2210) along with all the mentioned variables to have them ready to be analyzed.

2.5.1. Diversity metrics calculation

From the starting data, many metrics and indices were calculated, with the goal of obtaining robust data to characterize fish diversity, also decomposing the information for the specific diversity of each habitat, and their relative contribution. In particular, using the 'vegan' package in R, the species richness was computed for each transect. This is a basic diversity metric that reflects the number of different species in each replicate, giving an accurate estimate of the taxonomic diversity. Fish biomass (i.e. wet weight, in g) was estimated from size data by means of length-weight relationships from the available literature, selecting coefficients referring to Mediterranean samples whenever possible (Bayle-Sempere et al., 2002, www.fishbase.org). The biomass for each individual in each transect was then summed, to obtain the total biomass of each transect (community biomass, Cardinale et al., 2013) and also the total biomass of each species for other analytical procedures. Biomass, along with the abundance of every species in a transect, was calculated both for each site and also for each singular species within the sites, to have complete information on the presence and density of the fish species. Shannon-Wiener index and Simpson's index of Diversity were also computed by using the 'vegan'

package. Shannon-Wiener index measures the evenness of species in a community, meaning how similar the abundances of different species are in a given community. Its value rises with the number of species and the evenness of their abundance, and it is calculated as follows:

Shannon Index (H') =
$$\sum_{i=1}^{s} p_i \ln p_i$$

Where " p_i " is the frequency of the ith found on the total number of organisms in a sample (n_i / N), ln is the natural log, and "s" is the number of species.

The Simpson's Index of Diversity is a dominance index, because it gives more weight to common (or dominant) species, and it is calculated as:

Simpson's Index of Diversity (D) =
$$1 - \sum_{i=1}^{s} p_i^2$$

Where " p_i " is the proportional abundance of one particular species "ith" found, and "s" is the number of species.

This index represents the probability that two individuals randomly selected from a sample will belong to different species. Following this definition, the greater the value is, the greater will the diversity be.

All the metrics were calculated for each replicate (transect), to then be possibly pooled per habitat type (Rocky, Sandy, *Posidonia*) when specifically analyzing

the differences between the habitats or kept separate when there was a need to have the information in an extended way.

Finally, the total density (total number of individuals recorded within each transect) was calculated to have an overview of the mean number of fish counted over the transects, and eventually the number of "null transects" (i.e. transects without any observation).

Another part of the study focused more on the metrics for each separate species. To do so, the abundances and biomasses of each species in every transect were considered. The same procedure was also performed for the subsets that just include the observations from one habitat, to have a more straightforward information about the occurrence and density of species in every habitat.

Furthermore, the study aimed at finding any differences and uniqueness of the considered habitats. To do so, the "**dominant habitat**", intended as the habitat in which the abundance of a species was the highest, was identified. This value can give useful information about the habitat preference of a species and has been done also following Harper et al. (2022). To highlight this concept even more, also the species that were found in just one habitat, named "**unique species**", and the percentage of them on the total number of species found in that habitat, were detected (Harper et al., 2022). On the other hand, to assess potential similarity between the habitats, the species in common to two or all

of them were identified.

To find the species that have the highest "probability" of being observed associated in the same replicate, a **ranked crossed-correlation** was compiled and then plotted for the 20 most relevant "couples of species" (p < 0.05). All these results have been graphically visualized using the 'ggplot2' package in RStudio.

2.5.2. β -diversity estimates

Following this path, started by considering the α -components of diversity, intended as the local "richness", the **β-diversity** was evaluated, following the approach by Harper et al. 2022. β-diversity is intended in this study as "the extent of change in community composition" (Whittaker, 1960), in this case between the different habitat types. The β-diversity provides useful information about the contribution of each habitat in shaping the fish biodiversity and highlights the differences between them. β-diversity estimates were computed using the R package 'adespatial', which contains the function 'beta.div'. This function automatically calculates the two components of β-diversity of interest, based on the proposed division of the total β-diversity by Legendre and Cáceres (2013): Local Contribution to Beta Diversity (LCBD), that quantifies the ecological uniqueness of each site in relation to a wider ecosystem network (Heino and Grönroos, 2017), and Species Contribution to Beta Diversity

(SCBD), considered as the relative importance of each species in affecting beta diversity patterns (da Silva et al., 2018). LCBD is an index for the "community" uniqueness" of each site compared to the entire seascape (Legendre and de Cáceres, 2013). A high LCBD is indicative of the high dissimilarity between two habitats. This decomposition of the total change in community composition well fits one of the goal of the study, which is the evaluation of the habitats' contribution to diversity. In this case the LCBD values were plotted by grouping them per habitat type, just considering the significant values (p < p0.05), calculated following the Holm's method (Holm, 1979). Regarding the species, high SCBD values for one species indicate that a species highly differs in its distribution between the habitats of the study area (Legendre and de Cáceres, 2013). We then plotted the 10 species with the highest SCBD in ggplot2. The initial data were scaled using "Hellinger transformation" to obtain more suitable values for the analyses.

2.5.3. Analysis of variance (ANOVA)

To test putative difference in univariate diversity metrics (see above) between habitats and accounting for our experimental design (including locations and sites), we used the analysis of variance (**ANOVA**, package 'nlme' in R), designed with a mixed-effects model, in which the sites (62 levels) were nested in the locations (five levels), being both random factors, and being nested in the habitat (fixed, with 3 levels: rocky, sandy, *Posidonia*). This design was chosen to assess variability among habitats, accounting for spatial replication at the two spatial scales considered. To assess whether the inclusion of the nested factors improved the ANOVA or not, also models where each of the factors was gradually removed were performed, and the best model has been chosen looking at the AIC results. Post-hoc tests on the main factor (habitat) were performed using pairwise comparisons among the groups through "emmeans" package in R.

2.5.4. Permutational Multivariate Analysis of Variance (PERMANOVA)

To analyze the differences in multivariate densities between habitats, a **PERMANOVA** (Anderson, 2001) was carried out. This method belongs to the family of the "ANOVA-like tests" and constructs an output from a Bray-Curtis resemblance matrix calculated among the sample units and obtain "p-values" using random permutations of observations among the groups. The same experimental design used for ANOVA was adopted. Before running the PERMANOVA, data were square root-transformed, followed by the addition of a "dummy value", to tackle the problems regarding the many zeros in the data matrix, which is a common issue in ecological data (Zuur et al., 2010). The PERMANOVA analyses have been computed using PRIMER software, version 7.

2.5.5. Nonmetric Multidimensional Scaling (nMDS)

To graphically represent the multivariate information about the diversity in species composition and highlight the features of the different sites belonging to different habitats, a **Nonmetric Multidimensional Scaling** (nMDS) was carried out. This technique allows to represent the original position of communities in multidimensional space as accurately as possible using a reduced number of dimensions, using the rank orders rather than the absolute abundances (distances) and can be applied with every dissimilarity matrix (in this case using Bray-Curtis dissimilarity). The matrices were scaled using "Hellinger transformation", to avoid excessive effects related to very large values. Also in this case, a "dummy value" was added to the original data for computational reasons. To perform the nMDS in R, the 'metaMDS' function of the 'vegan' package was used.

2.5.6. Species Distribution Model (SDM)

The last step to analyze the obtained data about diversity was to relate it to the pressures and the environmental variables obtained as described before and geographically referenced for each spatial unit, to see how these variables shape fish diversity in the study area, and to make projections regarding the distribution of the considered fish species along the entire Calabrian coastal waters, possibly highlighting areas with high or low species richness

(respectively "hotspots" and "coldspots"). This was done by developing a **Species Distribution Model (SDM)**, a common tool in quantitative ecology (Thuiller et al., 2009), In our case, the SDM was implemented by using the specific richness of each transect, in order to relate the richness to the several drivers into account. Moreover, the general nature of this metric was used to predict the richness of the entire Calabrian coasts, always considering the spatial behavior of the drivers obtained from the CMEMS database and the ones from the field sampling. The drivers used were all the ones described in the paragraph 2.4., and the habitat type (categorical), that was highly relevant for our study purposes.

Firstly, all the spatial data concerning the possible drivers to investigate were gathered to check on the collinearity between them (Zuur et al., 2010), that can possibly result in a reduced performance and precision of the model, because of the "masking" of the real effect of one variable. The variance-inflation factor (VIF, Allison, 1999), which is a measure for the increase of the variance of the parameter estimates if an additional variable is added to the linear regression, was used for this purpose, and only the variables with a VIF value lower than 4 have been retained for further analyses. After this preliminary "filter", 3 different models have been fitted to our data: a linear model, a Generalized Linear Model (GLM), and an Additive Linear Model (GAM), with the Poisson

error distribution type. The three models were tested based on the Akaike information criterion (AIC) results and the graphic inspection of the residuals to select the best one.

Moreover, we used a backward selection approach, useful to fit the regression models in which the choice of predictive variables was carried out manually. To this end, the stepwise model selection was performed by comparing the AIC of the full model and the ones of all the possible combinations of reduced models, to finally select the model with the lowest AIC value and the lowest number of variables. Finally, an ANOVA on the terms selected was performed to check on their significance.

To predict the richness of the entire Calabrian coasts, the results of the model developed beforehand were extended, by using the georeferenced rasters regarding the area, obtained as previously mentioned from the CMEMS database (<u>http://marine.copernicus.eu</u>), and mapped on the cartography of interest. Also the errors associated to the model were computed and mapped to see whether their contribution could influence the results of the model itself.

3. RESULTS

Considering the total number of sites sampled (62), divided into 24 rocky sites, 23 sandy sites, and 15 *Posidonia* sites, 58 of them belong to the 0 - 20 m stratum(surficial), while 4 of them to the 20 - 30 m stratum (intermediate). The average depth of the sites was assessed at around 9.77 m, obtained by averaging the initial and the final depth recorded for each transect at each site.

First of all, a brief revision of the number of observations for each transect has been plotted (figure 3.1). In 8.5% of the total transects (42), no individuals were recorded (all the 42 null transects belong to sandy habitat replicates), and in 28% of them (138), the number of observed fishes did not exceed 5 individuals (including the null ones). On the other hand, 138 transects recorded more than 100 individuals, and 220 transects (44%) recorded a number between 6 and 100 fishes.



Figure 3.3. Number of fish observed for each transect (replicate) during samplings.

To focus on the high number of null transects, these data were divided among the habitats, explaining that almost all of them belong to sandy transects, which was the case during the sampling operations. The very high abundances recorded for some transects were mainly due to big schools of small fishes (e.g. *C. chromis*).

3.1. Species number

In total, 72 species were recorded, belonging to 25 families (Table 3.1). The most represented families are Labridae (13 species), Sparidae (12 species), Gobiidae (10 species), and Serranidae (7 species). Some of the species were regularly observed in a relevant number of replicates (e.g. *C. julis,* 52.6 % of the transects), while others were observed on just a few occasions (e.g. the family *Tripterygiidae*, 3% of the transects). In a very limited number of cases, fishes were identified at genus (e.g. *Trachurus, Sphyraena*) or family (e.g. *Mugilidae*) level due to the limitations of visual identification.

Table 3.1. List of the species found during the UVC sampling. Also the habitat in which each species has been most commonly found (dominant habitat), is noted (column three).

Family	Species	Dominant habitat	
Apogonidae	Apogon imberbis	Rocky	
Atherinidae	Atherina spp.	Posidonia	
Blennidae	Parablennius rouxi	Rocky	
	Parablennius tentacularis	Sandy	
	Parablennius zvonimiri	Rocky	
Bothidae	Bothus podas	Sandy	
Callionymidae	Callionymus spp.	Sandy	
Carangidae	Trachurus spp.	Sandy	
Centracanthidae	Spicara maena	Posidonia	
	Spicara smaris	Sandy	
Dactylopteridae	Dactylopterus volitans	Sandy	
Dasyatidae	Dasyatis pastinaca	Sandy	
Gobiidae	Deltentosteus quadrimaculatus	Sandy	
	Gobius bucchichi	Rocky	
	Gobius cobitis	Rocky	
	Gobius fallax	Rocky	
	Gobius geniporus	Rocky	
	Gobius incognitus	Sandy	
	Gobius niger	Sandy	
	Gobius xanthocephalus	Rocky	
	Pomatoschistus sp.	Sandy	
	Zebrus zebrus	Rocky	
Labridae	Coris julis	Rocky	
	Labrus merula	Posidonia	
	Labrus viridis	Rocky	
	Symphodus cinereus	Rocky	
	Symphodus doderleini	Rocky	
	Symphodus mediterraneus	Rocky	
	Symphodus melanocercus	Rocky	
	Symphodus ocellatus	Rocky	
	Symphodus roissali	Rocky	
	Symphodus rostratus	Posidonia	
	Symphodus tinca	Rocky	
	Thalassoma pavo	Rocky	

	Xyrichtys novacula	Sandy	
Mugilidae	Mugilidae	Rocky	
Mullidae	Mullus surmuletus	Rocky	
Muraenidae	Muraena helena	Rocky	
Pomacentridae	Chromis chromis	Rocky	
Scaridae	Sparisoma cretense	Rocky	
Scophthalmidae	Lepidorhombus boscii	Sandy	
Scorpaenidae	Scorpaena maderensis	Rocky	
	Scorpaena notata	Rocky	
	Scorpaena porcus	Rocky	
	Scorpaena scrofa	Posidonia	
Serranidae	Anthias anthias	Rocky	
	Epinephelus aeneus	Sandy	
	Epinephelus costae	Rocky	
	Epinephelus marginatus	Rocky	
	Serranus cabrilla	Posidonia	
	Serranus hepatus	Sandy	
	Serranus scriba	Rocky	
Sparidae	Boops boops	Rocky	
	Dentex dentex	Rocky	
	Diplodus annularis	Posidonia	
	Diplodus puntazzo	Rocky	
	Diplodus sargus	Rocky	
	Diplodus vulgaris	Rocky	
	Lithognathus mormyrus	Sandy	
	Oblada melanura	Posidonia	
	Pagellus acarne	Posidonia	
	Pagrus pagrus	Sandy	
	Sarpa salpa	Rocky	
	Sparus aurata	Posidonia	
	Spondyliosoma cantharus	Posidonia	
Sphyraenidae	Sphyraena sp.	Posidonia	
Syngnathidae	Hippocampus hippocampus	Sandy	
Trachinidae	Trachinus araneus	Sandy	
	Trachinus draco	Sandy	
Tripterygiidae	Tripterygion delaisi	Rocky	
	Tripterygion melanurus	Rocky	
	Tripterygion tripteronotus	Rocky	
Total families = 25	Total species = 72		

As we can notice in Table 3.1, it is possible to find (column 3) the "dominant habitat" of each species. This parameter has been evaluated by calculating in which one of the three habitats the abundance of a species was the highest. In general, a total of 56 species were found in rocky habitats, 38 species in *Posidonia* meadows and 32 species in sandy bottoms. These numbers do not exclude the co-occurrence of one species in more than one of the habitats, as these aspects of "uniqueness" will be presented later on.

The total number of fish individuals (without distinctions for the species) found was 30,955 individuals for the rocky bottoms, 10,177 for *Posidonia* meadows, and 2,027 individuals for sandy bottoms, highlighting evident differences in the magnitude of fishes observed in each habitat.

3.1.1. The abundances

Analyzing the species separately, some of them were found at high **densities** (mean \pm SE) most of the times they have been encountered. This is the case of species like *Chromis chromis* (44.8 \pm 4.19 individuals/125 m²), *Boops boops* (8.2 \pm 1.6 individuals/125 m²), and *Oblada melanura* (6.2 \pm 1.1 individuals/125 m²) (those data are referred to the observations on the 3 habitats pooled). Those species tipically form big schools. More in detail, in rocky habitats (figure 3.2a) the three species with the highest mean density are *C. chromis* (93.5 \pm 9.4 individuals/125 m²), *B. boops* (9.8 \pm 2.8 individuals/125 m²), and S. sarpa (9.4

 \pm 1.3 individuals/125 m²).

Posidonia meadows (figure 3.2b) showed similar trends in the densities of species, with the maximum values for *C. chromis* $(35.1 \pm 4.2 \text{ individuals}/125 \text{ m}^2)$, *B. boops* $(14.9 \pm 4.1 \text{ individuals}/125 \text{ m}^2)$, and Oblada melanura $(9.9 \pm 2.3 \text{ individuals}/125 \text{ m}^2)$.



Figure 3.4. Mean number of individuals (abundance) for each species in rocky (a), Posidonia (b), and sandy (c) habitats.

Lastly, Sandy sites (figure 3.2c) deviated a bit in their most abundant species, represented by *Oblada melanura* $(2.3 \pm 2.17 \text{ individuals}/125 \text{ m}^2)$, *B. boops* $(2.2 \pm 1.6 \text{ individuals}/125 \text{ m}^2)$, and *Xyrichtys novacula* $(2 \pm 0.23 \text{ individuals}/125 \text{ m}^2)$.

3.1.2. Fish biomass

Also fish **biomass** (fish net weight) was estimated (figure 3.3) (mean \pm SE), to have a better idea of the actual quantity of organisms, and in this case, other organisms have been highlighted for their biomass, such as *Sarpa salpa* (770 \pm 132.1 g/125 m²), *Diplodus vulgaris* (530.5 \pm 74.9 g/125 m²) and *Chromis chromis* (410.8 \pm 58.01 g/125 m²) on rocky substrates, *Oblada melanura* (70.9 \pm 68.8 g/125 m²), *Xyrichtys novacula* (40.28 \pm 5.7 g/125 m²), *Boops boops* (28.1 \pm 24.6 g/125 m²) on sandy substrates, and *Chromis chromis* (109.5 \pm 16.5 g/125 m²), *Oblada melanura* (86.1 \pm 33.9 g/125 m²), and *Sarpa salpa* (78.4 \pm 26.2 g/125 m²) in *Posidonia* meadows.



Figure 5.3. Mean biomass for each species in rocky (a), and sandy Posidonia (b), and sandy (c) habitats.

3.1.3. Species co-occurrence

To further understand the patterns of occurrence of the species and better explore the dataset, a ranked cross-correlation output was compiled (Figure 3.4). Among the five highest correlated couples of species, we can find the Family *Mugilidae* and *Scorpaena notata* (0.59), *Trachinus araneus and T. draco* (0.50), *Diplodus puntazzo* and *D. sargus* (0.43), *Parablennius zvonimiri*

and *Symphodus doderleini* (0.43), *Diplodus sargus* and *Sparisoma cretense* (0.42). Those are just the highest correlated examples of significative correlations (p < 0.05), that can inform about the "probability" of observing the two species considered at the same time (i.e. in the same replicate).



Correlations with p-value < 0.05

Figure 3.4. Ranked cross-correlations for the abundance matrix of the species found. The plot shows the 20 most correlated species pairs.

3.2. The habitats

3.2.1. Species richness per habitat

In terms of species richness (figure 3.5a), this had a mean value of 6.96 (SE \pm 0.2) species/125 m² in *Posidonia* meadows, mean number of 10.22 (\pm 0.19) species/125 m² was instead observed in rocky bottoms, while an average of just 1.26 (\pm 0.08) species/125 m² was found on sandy bottoms. Variability among

the groups was statistically significant (df: 6, F: 360.2, p < 0.05, ANOVA). The following pairwise comparisons for all the habitat pairs resulted significant as well. The addition of the nested and random factors (locations nested in habitat, and sites nested in the locations) gave better results (in terms of AIC values), and so it has been kept with the consideration of all the factors in the final model.

3.2.2. Diversity indices

The diversity indices provided a complementary piece of information for the diversity estimates of the habitat. The Shannon index (*H'*), representing the evenness component of a community along with the simple specific diversity, has given the highest values in rocky bottoms, specifically with a mean value of 1.43 (\pm 0.04), followed by *Posidonia* meadows, where the mean value of *H'* is 1.14 (\pm 0.04). Sandy bottoms showed instead very low values of the Shannon Index, stopping at 0.23 (\pm 0.03). The differences resulted statistically significant (df: 6, F: 166.9, p < 0.05, ANOVA) and the pairwise differences for the habitats assessed that each habitat was significantly different from each other. Shannon index showed a significative improvement in the ANOVA output when considering the nested random factors (localities and sites).

The results regarding the Simpson's Index showed similar trend, as the obtained results were 0.62 (\pm 0.015) for rocky bottoms, 0.5 (\pm 0.016) for

Posidonia meadows, and 0.37 (\pm 0.03) for sandy substrates. The differences between the groups were statistically significant (df: 6, F:11.6, p < 0.05, ANOVA) but in this case, the difference in the mean values between the rocky and *Posidonia* meadows were not statistically significant (assessed through pairwise comparisons). The inclusion of the nested and random factors (locations nested in habitat, and sites nested in the locations) improved the model also in this case.

3.2.3. Biomass per habitat

Biomass values (figure 3.5b) reflected the richness trends, with rocky bottoms showing the highest biomass (3949.8 ± 344.2 g/125 m²), followed by *Posidonia* meadows (738.7 ± 72.2 g/125 m²) and Sandy bottoms (244.3 ± 105.2 g/125 m²). The differences among the habitats were statistically significant (df: 6, F: 22.8, p < 0.05, ANOVA), but in this case, only rocky bottoms significantly differed from the other habitats (assessed through pairwise comparisons), while Sandy and *Posidonia* meadows did not show significant differences. The outcome of the ANOVA regarding the two random factors nested in the "habitat" improved the model also in this case.



Figure 3.5. Mean richness (a) and biomass (b) for each habitat.

3.2.4. Similarities and differences in species composition between the habitats In rocky bottoms, the number of unique species found was 16, followed by 14 unique species in sandy bottoms. *Posidonia* meadows did not record any unique species, which is quite an interesting outcome. Even more interesting was the percentage of unique species in relation to the total number of species recorded in each habitat. In fact, in sandy bottoms almost half (43.8%) of the total species are unique for this habitat, "outscoring" this value calculated for rocky habitats (28.6%).

On the other hand, the similarity between the assemblages in the different habitats was calculated (Figure 3.6). These values are in accordance with the previous results. In fact, the two habitat types sharing more species are rocky and *Posidonia* (24 species), while sandy habitats share not so many species

with the other two (4 species with rocky habitats and 2 with *Posidonia* ones). Furthermore, 12 species are shared between all the three considered habitats, such as *C. chromis, S. salpa* and *M. surmuletus*, possibly underlining their generalist habits.



Figure 3.6. percentage of species found in common between the habitats.

3.2.5. The β -diversity

β-diversity estimates have been divided in our case in the Local Contribution to Beta Diversity (LCBD) and Species Contribution to Beta Diversity (SCBD). The results regarding LCBD gave emphasis to the significance of the previous results, as they highlight a marked difference between sandy sites, being by far the habitat with the highest mean contribution to the LCBD (0.0025 ± 7.45×10⁵), as we can see in figure 3.7, meaning that the fish assemblages inhabiting this kind of habitat are confirmed to be the most "unique", while *Posidonia* and rocky bottoms have been showing lower mean values, respectively $0.0019 \pm 3.54 \times 10^{-5}$ and $0.0016 \pm 2 \times 10^{-5}$.



Figure 3.7. Local Contribution to Beta Diversity per habitat.

Regarding β -diversity, the ANOVA assessed that the differences between the groups were statistically significant (df: 6, F:27.6, p < 0.05), but the pairwise comparisons revealed that *Posidonia* and rocky habitats did not significantly differ. The best model for the LCBD component was again the "complete" one, that included both nested random factors.

To go deeper into the characterization of the β -diversity component of the Calabrian coastline, the SCBD was calculated (figure 3.8), to better understand which are specifically the species most responsible for the differentiation patterns identified for the habitats. The most influential species, with a very high SCBD value (0.18), is *X. novacula*, which has been already mentioned

before as one of the species inhabiting exclusively sandy bottoms, followed by *C. chromis* (0.12), *C. julis* (0.052), *O. melanura* (0.049), and *B. podas* (0.048).



Figure 3.6. Species Contribution to Beta Diversity. The colors of the column represent the dominant (or preferred) habitat for each selected species.

3.2.6. Assemblage composition in the three habitats

The graphical results of the NMDS (Figure 3.9), also completed by the calculation of clusters to highlight the habitat types, provided a good summary of the differences in assemblage composition between the sites, and their link with the species.

First of all, the calculation made by choosing a 2-dimensional output (NMDS1 and NMDS2), generated a stress value of 0.17, which states a good representation of the simplified community. The stress represents the difference between the data point position in the two-dimensional plot and the "real"

distance calculations in the multidimensional space. Moreover, the sites belonging to rocky bottoms did not show so much variability and even represented a sort of subset of *Posidonia meadows*, despite actually being the most diverse habitat if considering the diversity metrics. Posidonia sites shows a slightly higher variability, overlapping in few cases with the cluster of the sandy sites. Sandy bottoms are instead completely separated from the other two clusters, and the differences in the rank-based distances between the sites are more evident than in the other cases, forming a very wide cluster. The rankbased positions of the species, indicating how the species are distributed in the plot and how they link with the sites, are in accordance with the previous results, especially looking at the highlighted species, which are the ones with the highest values of SCBD, with X. novacula and B. podas being the most "distant" species from the rocky and *Posidonia* meadows, and on the opposite side C. chromis and O. melanura. More in general terms, we can say that the position of species in the plot overlapped quite well with their previously obtained preferred (or dominant) habitat.



Figure 3.9. Nonmetric Multidimensional Scaling for the sites (points) aggregated per habitat type (colors). The groupwise centroids (triangles) have been calculated to allow the clusterization and highlight the distinctions between the habitats. The species displayed in the labels (10) are the ones with the highest contribution to β-diversity.

The PERMANOVA (Table 3.2) on the abundance data matrix was useful as a confirmation of the graphical hints given by the NMDS. Its results revealed that the differences between the habitats for the fish assemblages were significant. Both location (df: 10, pseudo-F: 2.4, p < 0.05) and site (df: 49, pseudo-F: 4.5, p < 0.05) factors were significant.

The pairwise tests performed for the habitat factor confirmed that each pair of habitat tested was significantly different from each other (p < 0.05).

Table 3.2. PERMANOVA output on the square root transformed data of the fish abundance. (Significance : p < 0.05)

Source	df	SS	Pseudo-F	p (perm)
habitat	2	5.17 x 10 ⁵	25.504	0.0001
location(habitat)	10	1.14 x 10 ⁵	2.4339	0.0001
<pre>site(location(habitat))</pre>	49	2.31 x 10 ⁵	4.4813	0.0001
Residuals	434	4.56 x 10 ⁵		
Total	495	$1.42 \ge 10^6$		

3.2.7. Drivers of species richness and predicted patterns along Calabrian coasts

Among the three models' families tested at the beginning, the one with the best fit, based on the AIC results (table 3.3a), was the GAM-based model. The backward selection of the variables optimized the previously selected model with a lower AIC (1915.221), and finally allowed the composition of the final model, that is a GAM model with a Poisson error distribution composed of:

Richness ~ factor(habitat) + s(SST, bs = "cs") + s(SSS, bs = "cs") + chlWhere "habitat" is the main factor related to richness, "SST" and "SSS" are included in a smoothing term with a shrinked cubic regression spline, and "chl" is linearly correlated to the response variable. The results of the ANOVA (table 3.3b) regarding the variables included in the model resulted significative for all the factors (p < 0.05) apart from "chl" (p = 0.0594), that was still included by the automatic computation of the model.

Table 3.3. (a) Results of the AIC, representing the model fit; (b) Results of the ANOVA on the terms of the chosen GAM model. The df values noted with "*" are referred to "expected" degrees of freedom.

a)	b)		b)	Factor	df	Chi squared	p-value
	Model	AIC		habitat	2	882.610	<2e-16
	Linear	2038		chl	1	3.553	0.0594
	GLM	1943		s(SST)	1.609*	7.452	0.00396
	GAM	1942		s(SSS)	5.587*	62.163	< 2e-16

The model results showed that the habitat factor (figure 3.10a) appears highly significant in shaping richness values, producing evident differences in the results, with sandy bottoms being the less rich in species, the rocky bottoms being in the opposite side the richest habitats, and *Posidonia* meadows assemblages being slightly less rich than rocky, but more than sandy assemblages. In regard to the environmental variables, fish richness shows an evident decline as the SST (figure 3.10b) and SSS increase (figure 3.10c). In particular, richness values decrease steeply as the temperature exceeds 21.2 °C.

As for the chlorophyll-a concentration (figure 3.10d), this variable causes similar responses to SST and SSS, so its increase leads to a reduction of species richness, but the related plot turns out linear, because the term included in the model is linearly correlated to the response variable.



Figure 3.70. Results of the GAM model for the specific richness regarding each individual parameter. (a) habitat, (b) SST, (c) SSS, and (d) chl.

Finally, the results obtained have been used to predict the possible patterns of species richness along the entire Calabrian coastline (figure 3.11).

The general trend concerning most of the extension of coastal habitats (< 30 m) is of a low species richness, around 3-4 species for every spatial unit (125 m²). Nonetheless, there are some areas with a higher number of species predicted by the model, and, especially around Vibo Valentia, there are some spots in which the predicted species richness is around 13-14 species, that is a major increase with respect to the rest of the coast, and it can be further defined as an "hotspot" for the Calabrian coastlines.

Another zone in which there is higher richness than average is located on the opposite side of Calabria, in particular in correspondence to the Marine Protected Area of Capo Rizzuto and surroundings, on the Ionian coast. In this case, the species richness is less than in the previous case, around 7-8 species per spatial unit predicted, but there are some smaller patches in which richness reaches values of about 11-12 species. The last zone that revealed interesting outcomes regarding the response variables is located in the Messina strait, where richness goes up to 12-13 species.

The error distribution has been graphically represented as well, and in general terms, the error is quite low (around 1-2 species), so it is confident to assess that the predictions are accurate in most of the cases.



Figure 3.11. Species Richness in the Calabrian coasts, based on the prediction of the Spatial Distribution Model. The smaller panel indicates the error distribution for the model.

4. DISCUSSION

The characterization of coastal fish diversity along Calabrian coasts resulted in a total of 72 species belonging to 25 families observed. This result is consistent and even slightly higher than other studies performed in the Mediterranean Sea using the same methodologies (UVC) and considering similar heterogeneity regarding the habitat-types sampled (Guidetti, 2000, Franco et al., 2006, La Mesa et al., 2011). Moreover, the results may represent a baseline for future comparisons, given the scarcity of such kind of studies (Guidetti, 2000, La Mesa et al., 2011), considering multiple habitats in coastal ecosystems (i.e. rocky bottoms, seagrasses meadows, and sandy bottoms), and the rapid changes the Mediterranean basin is experiencing. In fact, other investigations only focused on rocky fish assemblages (De Girolamo and Mazzoldi, 2001, Piazzi et al., 2012, Carminatto et al., 2020, Bevilacqua et al., 2021), or seagrasses meadows (Bell and Harmelin-Vivien, 1982, Moranta et al., 2006, Deudero et al., 2008), but the addition of sandy bottoms resulted in an increase of the species encountered, due to their contribution to the total pool. In fact, a maximum of 65 species were recorded in the other studies (Piazzi et. al., 2012, focused on rocky reefs assemblages). This number results even higher than ours if we just consider rocky bottoms, but the consideration of a complete landscape composed by more than just one specific habitat, as in our case, increases the
total number of species. Moreover, the habitat factor was never considered in other studies (Lazzari et al., 2020).

4.1. Overall species abundance and biomass of the Calabrian coasts

To go more in details on the taxa found, the families that were represented by more species are Labridae (13 species), Sparidae (12 species), Gobiidae (10 species), and Serranidae (7 species). These families are generally well represented in Mediterranean coastal systems, as they include very heterogeneous species, capable of inhabiting all the habitat-types present in the shallow layers. For example, fish belonging to the *Labridae* observed during this study include species such as *Coris julis*, known to be widely spread all over the rocky habitats and in *Posidonia* meadows of the Mediterranean Sea (Fruciano et al., 2011), and others being more specialist regarding the habitat use. For example *Symphodus rostratus* is known to be commonly found in *Posidonia* meadows (Deudero et al., 2008), as also confirmed by our results, while others almost exclusively inhabit sandy bottoms, as in the case of *Xyrichtys novacula* (Alós et al., 2012).

Similarly, *Sparidae* family includes species from the genus *Diplodus*, commonly found in infralittoral rocky habitats (Sala and Ballesteros, 1997), and *Posidonia* meadows (i.e. *Diplodus annularis*, Francour, 1997) and other

species like *Lithognathus mormyrus* that occur predominantly over unvegetated sandy habitats (Guidetti, 2000, Matic-Skoko et al., 2007).

So, the wide-spectrum habitat-use for the organisms belonging to the mentioned families could be one of the reasons favoring their presence with high specific diversity over the study.

At species level, *Chromis chromis*, *Boops boops*, and *Oblada melanura*, are the most abundant species, usually occurring in large schools (Allam, 2003, Gkafas et al., 2013, Pinnegar, 2018). Their presence in the habitats sampled is quite consistent. Another abundant species was *Xyrichtys novacula* although this has been observed only in sandy habitats, in accordance with literature on its life habits (Alós et al., 2012).

Biomass measurements (intended as community biomass), that represent instead the fish net weight and could provide an indication about the stability of the community (Cardinale et al., 2013), highlighted a slightly different picture for the principal contributors to the biomass in the transects. This is the case of *Sarpa salpa* and *Diplodus vulgaris* for rocky substrates, that were recorded not only with multiple specimens for most of their observations, but can also reach quite large body-sizes (Gordoa et al., 1997, Buñuel et al., 2020), so they can easily be dominant in terms of biomass. For similar reasons, *Posidonia* meadows recorded highest biomasses values for *Oblada melanura*, that show comparable characteristics, but holds higher affinity with this habitat (Kalogirou et al., 2010), and for *Chromis chromis*, probably due to the very high densities recorded in this habitat. In sandy bottoms, the species showing the highest biomass values is *Xyrichtys novacula*, in accordance with the previous considerations about the abundance. In this case, this result could be due to its diffused presence in most of sandy replicates, but also, in some cases, because of the lack of other organisms. In fact, the dominant species recorded both for *Posidonia* and sandy habitats have a mean biomass one order of magnitude lower than the ones in rocky bottoms, and this difference is relevant and should be mentioned as we are considering the relative dominance of fish species for each habitat.

4.1.1. The Species Contribution to Beta Diversity (SCBD)

Here we follow the apporach to β -diversity estimations suggested by Legendre and de Cáceres (2013), so both the species contribution (SCBD) and the contribution of the habitat (LCBD) to β -diversity were considered crucial to combine the fish assemblages with the habitat in which they are hosted.

In fact, the first contributor to β -diversity among the species found is *X*. *novacula*, which is one of the species uniquely inhabiting sandy bottoms. This species inhabits the sandy and muddy substrates of almost all the western Mediterranean Sea (Castriota et al., 2005). It is reported that *X. novacula*

mainly feeds on small organisms, mostly mollusks, but also on crabs, shrimps, polychaetes and other inhabitants of soft bottoms (Cardinale et al., 1997). The species spends its whole life in such substrates with an haremic social organization (Candi et al., 2004), which is typical of protogynous species (Plant and Zeleznik, 2014) like *X. novacula*. Despite of its quite generalist habits, this species seems to not compete with other species of sandy habitats, likely because of its high capacity to survive in open sandy environment, by burying itself under the sand in presence of possible threats (Alós et al., 2012).

Chromis chromis, which is the second contributing species to β -diversity, is numerically the most abundant in the whole study area, partially explaining this high SCBD value (Carlucci et al., 2018). In fact, this species is very common in the coastal Mediterranean basin (Pinnegar, 2018), mainly feeding on plankton, and so not strictly associated to the benthic compartment as most of the other species considered. So, it is not easy to discriminate if its presence in one habitat is just transient or due to the exploitation of the resources provided by the habitat itself. We need also to consider the high patchy distribution of the habitats in the sampling locations, although the abundance of the species recorded in sandy habitats is considerably lower (one order of magnitude) than that recorded in the other two habitats.

Similar considerations could be made for O. melanura, that has quite similar

life habits to *C. chromis*, showing prevalent schooling behavior and a relevant motility among different habitats (Gkafas et al., 2013). The species prefers *Posidonia* meadows although with similar abundances to rocky substrates, and even in sandy substrates it showed quite high density compared to the other species.

Coris julis is another generalist species, but it is more strictly associated to the benthic compartment than the previous ones, being more sedentary (Fasulo et al., 2010) and defined as omnivorous, with preference for animal material (Karachle and Stergiou, 2008). This labrid has been observed predominantly in rocky substrates, but also in *Posidonia* meadows, while just rarely found in sandy habitats, even if its possible relationship with soft substrates has been already documented in past studies (Tortonese, 1970).

Bothus podas is uniquely found in sandy habitats, in agreement with literature data (Abid et al. 2013), although it has also been found on seagrasses of the Mediterranean Sea (Schintu et al., 1994). The diet is similar to *X. novacula*, being mainly constituted by crustaceans, mollusks, bryozoans, annelids, and other invertebrates (Abid et al., 2013). Also its social strategy is almost identical, as it forms harems as well (Carvalho et al., 2003), but in this case the species is gonochoric. The similarities in the feeding and even more in the social-reproductive strategies, could be defined as some of the factors for their

"success" in driving β -diversity patterns, but many others should be investigated (Heino and Grönroos, 2017). Moreover, the density of *B. podas* is not as high as the other species that contribute the most to β -diversity, even if considering just the sandy habitats, so its SCBD value becomes even more relevant, especially in validating the result obtained.

Another interesting species contributing to β -diversity is *M. surmuletus*, also in relation to its interesting position in the nMDS output. In fact, despite the dominant habitat in which it has been observed is rocky bottoms, it seems its position is "in the middle" between the clusters of rocky/*Posidonia* and the sandy one, being even included in the latter. This species can be found both in hard and soft substrates, especially due to its foraging behavior (Tserpes et al., 2002; Uiblein, 2007), based on small benthic invertebrates (Labropoulou et al., 1997), thus leading to a marked potential for mobility across different habitats (Ajemian et al., 2016).

These results were quite informative in connecting the observations regarding the species and the overall contribution of the habitats in creating diversity patterns.

4.2. Fish diversity in the different habitats of Calabrian coasts

One of the main goals of this investigation was to assess the specific diversity

of each habitat.

Species richness statistically differ between habitats, and the highest values belong to rocky bottoms, followed by *Posidonia*, and lastly by sandy habitats. First of all, there is no extended information about how richness could variate in a landscape composed by all the habitats hereabove and considered in its completeness, at least when habitat is treated as the main factor of the analysis. In fact, proofs about the role of habitat complexity are most of the times limited to just one type of habitat, mainly rocky bottoms or seagrasses meadows (Staveley et al., 2017; Di Franco et al., 2021) or focused on tropical reef systems (Jenkins and Wheatley, 1998; Gratwicke and Speight, 2005; Ford et al., 2017), and few of them (Jenkins and Wheatley, 1998; Harper et al., 2022) covered a habitat variety large enough to give space to comparisons. In comparable investigations (Jenkins and Wheatley, 1998; Guidetti, 2000; Harper et al., 2022), more complex habitats, such as rocky bottoms (or reefs in the tropical environment) were richer in fish species, while Posidonia meadows, and even more sandy bottoms, showed lower complexity, highlighted as the first driver influencing fish assemblages (Hall and Kingsford, 2021). These results fit well with the ones obtained by our investigation.

The Shannon (H') and Simpson's (D) indices were useful to have an additional comparison with the richness values, giving their power to describe different

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properties of the community regarding fish diversity. The results confirmed that rocky bottoms are generally more diverse, showing higher values for rocky habitats both for H' and D, in both cases followed by *Posidonia* meadows and lastly by sandy bottoms. Despite of this, *Posidonia* and sandy habitats were not significantly different regarding the Simpson's index (p < 0.05). Rocky bottoms not only show higher richness, but they probably highlight a more uniform distribution of the inhabiting species, meaning that all the species found are generally more evenly distributed in rocky bottoms, while the distribution of the species in sandy habitats is less uniform. Moreover, the values regarding the Simpson index could be a proxy that some species (the dominant ones) play a greater role in shaping the diversity of rocky bottoms than in the other habitats but, in this case, the difference between *Posidonia* and sandy habitats is less evident.

Studies carried out in the Mediterranean basin about fish biomass did not find a clear relationship of it with habitat complexity (Di Franco et al., 2021), although the results were limited to rocky assemblages and other drivers in that case may be influential, like the protection level (Guidetti et al., 2014; McClure et al., 2020). Our results proved a marked difference based on the habitat type, with rocky bottoms clearly outscoring the values of biomass with respect to *Posidonia* meadows and sandy bottoms, being not significantly different from each other (p < 0.05). Thus, habitat complexity likely contributes to the difference in biomass found in each habitat, together with food availability (Anderson and Sabado, 1995) and many other factors (McClanahan et al., 2019).

An interesting property of biomass measurements is its power to estimate both the quantity of organisms, and partially avoid a too simplistic consideration of all the species and individuals as "equal entities". The difference between richness in rocky habitats and *Posidonia* meadows is less evident than the difference in their biomass. This difference could be due to the lower mean size of specimens from *Posidonia* meadows, acting as nursery areas and hosting the early life stages of many fish species, or because of small differences in the fish assemblages from the two habitats, as also suggested by the PERMANOVA results.

A different picture is outlined when looking at the fish communities in terms of unique species. The number of unique species, i.e., species recorded in just one habitat, provide information on how much each habitat contributes to the overall diversity (Hall and Kingsford, 2021). In this case, the pattern described so far is different, as rocky bottoms maintain the highest value in absolute terms (16 species), but followed by sandy habitats (14 species), with *Posidonia* meadows showing any unique species. The significance of this result is even more relevant if put in terms of relative number of unique species compared to the total species in the community. In this case, sandy habitats recorded 43.8 % unique species, while rocky habitats just 28.6 %. So, sandy assemblages already appear to show a more marked difference in species composition compared to the other habitats. The dissimilarity of *Posidonia* meadows and rocky bottoms with sandy ones is confirmed also by the number of common species: just four species with rocky habitats and two species with *Posidonia* meadows.

Looking at the graphical output of the nMDS, replicates belonging to *Posidonia* meadows and rocky bottoms show high similarity in their associated diversity, with the clusters formed by them being almost overlapped. In particular, rocky bottom seems to be a sort of sub-sample of *Posidonia*, and so the distinction in the species present in this two habitat is not so evident. This is instead the case of sandy bottoms, that segregated in the nMDS from the other two habitats. Furthermore, also the distance within the sandy replicates seems to be greater in the nMDS graph, likely due to higher heterogeneity of the composition of sandy bottoms' replicates. These results confirm the expected contribution of each habitat to the overall diversity of the area, which was one of the expected goals of the investigation.

4.2.1. The Local Contribution to Beta Diversity (LCBD)

An important step toward the process of estimating the contribution of each habitat to the overall diversity was the evaluation of β -diversity, as the division proposed by Legendre and de Cáceres (2013), i.e. LCBD and SCBD, that is the contribution of each habitat to the diversity (LCBD), and the identification of the species that contribute the most to dissimilarity among habitats (SCBD).

To our best knowledge, this approach has been rarely applied in the Mediterranean (Carlucci et al., 2018), and never as a tool coupled to the visual census sampling approach, as most of the times β -diversity is divided into "turnover" and "nestedness" components, meaning respectively the "replacement" of species and the "loss" of species between assemblages (Baselga, 2010). However, the potential and range of application of the proposed division of β -diversity is quite high (Harper et al., 2022).

In this specific case, the mean contribution of sandy habitats has been surprisingly pointed out as the most relevant, notwithstanding their underestimated importance as habitats contributing to the diversity of a seascape (Henseler et al., 2019). At the opposite end, rocky bottoms are the less relevant in terms of habitat contribution to β -diversity when considering this approach. This of course does not mean that sandy habitats are "more important" in providing diversity than the other two. As we know, also from

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the diversity metrics hereabove, the contribution of rocky habitats and, to a lesser extent of *Posidonia* meadows, is not neglectable, but it appears important to consider a seascape in its integrity, as also the "less diverse" sandy habitats are here proven to be an enrichment to the total diversity, probably because of the almost completely different species that can be found there.

In addition, when analyzing the species contributing the most to β -diversity (SCBD), among the six most contributing ones, two are uniquely present in sandy bottoms (*X. novacula* and *B. podas*), and the other four (*C. chromis, O. melanura, C. julis,* and *M. surmuletus*) are generalist species, found during the study in all three habitat types, with differences in their relative densities on a case-by-case basis.

4.2.2. Common species among the habitats and connectivity

The number of common species between the habitats should be considered as well, especially when evaluating a complex seascape as the one under analysis. In this case, the high number of species in common between rocky and *Posidonia* assemblages (24) could be a proxy of the organisms moving between the two habitats, possibly bringing the overall system to higher abundances and biomass (Vega Fernández et al., 2008). The connectivity between seagrasses and hard bottoms is generally acknowledged in coastal seascapes (Jaxion-Harm et al., 2012), as fish species exploit these habitats in different life-stages and

can transit between them (Hyndes et al., 2018; Díaz et al., 2019; French et al., 2021). Of course, every habitat still holds a certain degree of differentiation in the fish assemblages composition, especially regarding the quantitative measures (abundances) of the species, as also suggested by the results of the PERMANOVA, but *Posidonia* meadows did not show any "unique" species, and this could be a good indication of the connectivity between the two habitats. Sandy bottoms instead show a low number of species in common with the other two habitats, with a very typical fish assemblages, that probably are a major driver for the patterns of β -diversity. To go further in the investigation of the connectivity between the habitats, such as the tagging of some key fish species in marine assemblages that show a good degree of distribution in the coastal habitats (Di Lorenzo et al., 2016) in order to assess the real connection between the different habitats.

4.3. *Drivers of species richness and predicted patterns along Calabrian coasts* The habitat-type was the main driver here investigated, although other variables were identified as possibly influencing the patterns related to species richness in the Calabrian coasts. The Species Distribution Model (SDM) gave us the possibility to simultaneously underline how biotic and abiotic drivers could be related to the richness values, and all the relationships found were similar in their behavior. The significant variables obtained for the model into account (SST, SSS, chl-a; see Methods section, paragraph *2.5.6.* for extended information) are among the most important ones in influencing the biotic component of marine ecosystems. In particular, as the Sea Surface Temperature (SST) and Sea Surface Salinity (SSS) increased, the specific richness decreased. This can suggest inferences when considering the trends of these variables, especially SST, in the last decades for the Mediterranean Sea (Sakalli 2017), and the concerning projections for the ones to come (Shaltout and Omstedt, 2014), that can be influential factors in predicting how richness in this area could be shaped in the near future.

Furthermore, the increasing SST could lead to tropicalization events (Ben Rais Lasram et al., 2008), changing the dynamics of the ecosystems because of the appearance of Non-Indigenous Species (NIS), that already increased steeply in the last decades in the Mediterranean Sea (Azzurro, 2008). So, while temperature trends could positively affect the abundance of some species found in the area (e.g. *Sparisoma cretense*, Astruch et al., 2016), most of them could be affected in the opposite way. It is worth noting that, during the Visual Census activities, no NIS has been encountered by the operators in the study area.

The salinity (SSS) had negative effects on species richness as well, but evidence of this on a larger scale are not so investigated as in the previous case. Nonetheless, it is known that the trend of salinity is constantly increasing in the last decades, similarly to temperature (Jordà et al., 2017). The effects of salinity changes include a modification of the water masses circulation (Astraldi et al., 2002), with influences on the provision of food, nutrients and on the larval dispersion for fish species (Reiss et al., 2000; Williams and Follows, 2003; Tiedemann et al., 2018). In addition, salinity increase could be a factor (along with temperature) favoring the spread of NIS, especially the ones coming from the Red Sea (i.e. Lessepsian species), that will find suitable conditions for their settlement (Raitsos et al., 2010).

The relation found with chlorophyll levels is interesting as well, showing a linear reduction of species richness as the values increase. The effects of this factor are not always the same on an ecosystem and can be both related to an increase in richness values (Shah Esmaeili et al., 2022), but also be indicators of eutrophication phenomena after a certain threshold, bringing negative consequences to the ecosystem (Bell et al., 2014). So, while the effects in the range analyzed are negatively correlated with richness values, there are interesting results about how the increasing concentration of chlorophyll-a could be inversely related with the SST increase (Kotta and Kitsiou, 2019), leaving a high space for uncertainty regarding the relative importance of these drivers and their possible synergistic effects in shaping the fish assemblages of

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this area.

Biomass has not been directly related to the drivers in our case, but there is evidence of a possible influence of them to it (Duffy et al., 2016).

The second scope for the development of a Distribution Model for Species richness was a projection of the richness values on the entire Calabrian coastal waters. The results of the model confirmed the trend of the sampled locations, with a low specific richness spread throughout the majority of the coastline, probably due to the prevalence of long stretches of sandy bottoms in the coastlines, that were proven to be less rich than the more complex habitats of the coastal system of this area. Moreover, two areas have been highlighted as more diverse: (1) the Marine Protected Area (MPA) of Capo Rizzuto (and surroundings) and (2) the area around Vibo Valentia. Separate consideration must be made on the two cases. The first one is the only MPA of the Calabrian coastline, and the higher richness values predicted could have been among the criteria for the establishment of this protected area, as it is one of the most heterogeneous spot in the Ionian coastline, with many habitats such as rocky bottoms and *Posidonia* ecosystems within a relatively small area, and thus hosting higher specific richness.

In the second case instead, there are no measures applied at the moment in the area, and the touristic pressure is quite high compared to other locations.

Despite of this, fish biodiversity values showed a marked positive difference from the rest of the coastal waters, and this should push to consider this area as a possible candidate to be protected, also giving the requirements brought by the global 30 x 30 initiative, that aims at protecting 30% of the world's total lands and waters by 2030, with 10% strictly protected. Of course, a deeper comprehension of the potential of this area should be obtained by sound assessments that focus on multiple aspects of biodiversity (Mouillot et al., 2011), but this could be a good starting point to discuss about its future protection.

4.4. Biases and strengths of the study and other possible approaches

Looking at the depth range considered, the visual technique adopted, Underwater Visual Census (UVC), fits perfectly and complements other methods that could possibly be used for future assessments of the area, and already performed by other team members during the sampling campaign, but not analyzed in this particular investigation. Such methods are for example Baited Underwater Videos (BUVs), which could expand the operative range for the analyses at lower depths thanks to the absence of a diving operator, even if also still holding some limitations in their applicability, like the need to choose a priori a fixed spot where to deploy the instrument, without the flexible approach typical of human-based methods (UVC in this specific case), and the limited visual field, that can be partially solved by using stereo/360° cameras (Mallet et al., 2021).

Other possible methodologies involve molecular techniques, like the environmental DNA (eDNA), which represents the cumulative genetic material that can be obtained from natural releases in an environmental medium like water. This tool has been revealed very useful in ecological studies (Fraija-Fernández et al., 2020), thanks to its ability to capture a wide but somehow different spectrum of species, often just partially overlapping with the biodiversity resulting from the traditional survey methodologies (Kelly et al., 2017). DNA is then processed with PCR and next-generation sequencing methodologies to build "libraries" of the collected material.

Finally, new approaches to be considered rely on "new" technologies that are reducing the limitations related to "classical" diving techniques, such as the use of Closed Circuit Rebreathers (CCR). This technique not only allows a partial overcoming of the limits related to depth thanks to their efficiency, and a stretch in the dive times with shortened decompressions, but also determines a less impacting presence of the operator, thanks to the avoidance of air bubbles production (Sieber and Pyle, 2010; Shepherd et al., 2021).

All these considerations led our investigation toward this methodological

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direction.

Of course, the UVC holds some limitations as well, like the biases related to the experience of the observer and some possible behavioral changes in response to the diver presence (Kulbicki, 1998; Dickens et al., 2011), but it returns fast and quite accurate estimates while containing the costs. Furthermore, in deeper waters, SCUBA diving using classical "recreative" gears become challenging and so does diving for scientific purposes (i.e. UVC), because of the limited time that could be safely spent at such depths and the eventual need to perform complex diving profiles, requiring highly trained operators and specific equipment, and resulting in low replicability potential and long sampling times. To accomplish sampling at these depths, the use of video tools (i.e. BUVs) or other mentioned techniques could represent valuable alternatives as a visual approach.

Another portion of the study that could possibly be implemented is the lack of a temporal replication, for example in relation to seasonal variability. As widely known, seasonal patterns show high influence on the composition of fish assemblages (Claudet et al., 2006; Henriques et al., 2013), especially in temperate basins like the Mediterranean Sea, where oscillations in the abiotic, and consequently biotic factors (Duarte et al., 1999; Trabelsi and Rassoulzadegan, 2011) could play a relevant role in the marine ecosystems, worth to be analyzed more in detail.

It is also important to mention that the methods used during the sampling procedures do not rely on catch-dependent data and so, while they still have all the above-mentioned limitations, they avoid all biases concerning the specific gears used for sampling in catch-dependent approaches (i.e. nets, vessels, etc.), which could lead to a possible misleading community composition due to a reduced range of taxa, trophic levels and size classes considered (Caldwell et al., 2016). So, while sampling did not just focus on fishery-targeted organisms but on all the species detected, allowing a more complete understanding of the overall ecological effects, also in relation to the habitat under the study lens, the results have the potential to improve many sectors, fisheries included, that could benefit from an enhancement of the accessible information, proceeding towards a more responsible approach to the sustainable resource management. The key to increase the possibilities of a reliable and robust assessment is to increase the amount of available data for the area, to allow the adoption of multiple approaches using all those mentioned techniques in the most efficient way possible, avoiding an excessive data overlap but having the possibility to obtain a complete assessment on the topic. Moreover, no method is more correct than others, and the most important features that must be considered are complementarity and intercomparability (Aglieri et al., 2021), achievable only

with the use of standardized procedures during the sampling design (Caldwell et al., 2016). This in the scope of a possible creation of timely and spatially extensive datasets and increase the knowledge about the fish diversity, to increase the reliability of the estimates and provide a baseline for any future actions aiming at protecting the biodiversity of the area. This planning should also consider how the different habitats can contribute to create a diverse seascape, with the inclusion of the often underestimated sandy habitats, that were proven to enhance the overall regional diversity thanks to unique fish assemblages they host and their unexpected contribution to β -diversity.

4.5. Conclusions

The overall aim of this study was the assessment of coastal fish diversity and its drivers in the different coastal habitats. Following the three main goals, it is possible to conclude that:

1. The assessment of the **fish diversity of each habitat** under consideration resulted in a higher diversity of rocky habitats in terms of species richness, supported by the results regarding the biomass, and the diversity indices (*H*' and *D*). Sandy bottoms were instead the less diverse habitat in relation to all these metrics, while *Posidonia*

meadows held an intermediate position, although some metrics, especially regarding biomass, were really low as well.

- 2. The **contribution of each habitat** to the overall diversity, achieved through the analysis of β -diversity, highlighted instead a high contribution of sandy bottoms to the degree of difference between the habitat-specific assemblages, determining the high importance of sandy assemblages in the creation of separated fish assemblages. In this case, the trends of α -diversity were reverted, with rocky bottoms showing the lowest values of the two components in which β diversity was divided (LCBD and SCBD). Posidonia assemblages were again in an intermediate situation, but other results such as the lack of unique species allows to infer about their hypothetical connectivity with rocky habitats and explain their low contribution to β -diversity. These results could be useful in future assessments, also in relation to the inclusion of a wider spectrum of habitats in marine spatial planning initiatives.
- 3. The **Upscaling** of the **biodiversity assessment**, accomplished with the use of a Species Distribution Model focused on species richness, provided interesting results on the decrease of the richness values with the increase of the variables considered (SST, SSS, and Chl-a),

indicating a possible relation with the present and future trajectories of climate change. Moreover, the predictions on the whole Calabrian coasts highlighted two areas with higher richness values: the area around the MPA of Capo Rizzuto (on the Ionian side), which is already protected to preserve its biodiversity, and the area around Vibo Valentia, that shows even higher richness values, that could be a priority area for protection in the future, towards the accomplishment of the 30x30 goal by 2030 (European commission, 2020; Mammola et al., 2020; Hermoso et al. 2022)

5. REFERENCES

- 1. Abid, S., Ouannes-Ghorbel, A., Jarboui, O. et al. (2013). Diet composition and feeding habits of the wide-eyed flounder, Bothus podas, in the Gulf of Gabes (Tunisia). Mar Biodiv 43, 149–161. https://doi.org/10.1007/s12526-012-0142-3
- Aglieri, G, Baillie, C, Mariani, S, et al. (2021) Environmental DNA effectively captures functional diversity of coastal fish communities. Mol Ecol. ; 30: 3127–3139. https://doi.org/10.1111/mec.15661
- Ajemian, M.J., Kenworthy, M.D., Sánchez-Lizaso, J.L. and Cebrian, J. (2016), Aggregation dynamics and foraging behaviour of striped red mullet Mullus surmuletus in the western Mediterranean Sea. J Fish Biol, 88: 2051-2059. https://doi.org/10.1111/jfb.12932
- 4. Alistair J. Cheal, Michael J. Emslie, Leanne M. Currey-Randall, Michelle R. Heupel, Comparability and complementarity of reef fish measures from underwater visual census (UVC) and baited remote underwater video stations (BRUVS), Journal of Environmental Management, Volume 289, 2021, 112375, ISSN 0301-4797, https://doi.org/10.1016/j.jenvman.2021.112375.
- ALLAM, S. (2003). Growth, Mortality And Yield Per Recruit Of Bogue, Boops Boops (L.), From The Egyptian Mediterranean Waters Off Alexandria. Mediterranean Marine Science, 4(1), 87–96. https://doi.org/10.12681/mms.244
- 6. Allison, P. D. (1999). Multiple Regression: A Primer. Thousand Oaks, CA: Pine Forge Press. p. 142.
- Alós J, Cabanellas-Reboredo M, Lowerre-Barbieri S (2012) Diel behaviour and habitat utilisation by the pearly razorfish during the spawning season. Mar Ecol Prog Ser 460:207-220. https://doi.org/10.3354/meps09755
- 8. Anderson M.J. (2001). Permutation tests for univariate or multivariate analysis of variance and regression. Canadian Journal of Fisheries and Aquatic Sciences. 58(3): 626-639. https://doi.org/10.1139/f01-004
- 9. Anderson, T. W., & Sabado, B. D. (1995). Correspondence between food availability and growth of a planktivorous temperate reef fish.

Journal of Experimental Marine Biology and Ecology, 189(1-2), 65-76. https://doi.org/10.1016/0022-0981(95)00011-F.

- 10. Araújo, M.B. and Luoto, M. (2007), The importance of biotic interactions for modelling species distributions under climate change. Global Ecology and Biogeography, 16: 743-753. https://doi.org/10.1111/j.1466-8238.2007.00359.x
- 11.Arndt, E.; Givan, O.; Edelist, D.; Sonin, O.; Belmaker, J. Shifts in Eastern Mediterranean Fish Communities: Abundance Changes, Trait Overlap, and Possible Competition between Native and Non-Native Species. Fishes 2018, 3, 19. https://doi.org/10.3390/fishes3020019
- 12. Astraldi, M., Gasparini, G., Vetrano, A., & Vignudelli, S. (2002). Hydrographic characteristics and interannual variability of water masses in the central Mediterranean: a sensitivity test for long-term changes in the Mediterranean Sea. Deep Sea Research Part I: Oceanographic Research Papers, 49(4), 661-680. https://doi.org/10.1016/S0967-0637(01)00059-0
- 13.Astruch, P., Bonhomme, P., Goujard, A., Rouanet, E., Boudouresque, C., Harmelin, J., & Harmelin-Vivien, M. (2016). Provence and Mediterranean warming: the parrotfish Sparisoma cretense is coming. Rapports de la Commission Internationale pour l'Exploration Scientifique de la Mer Méditerranée, 41, 362.
- 14. Azzurro, E., & Commission Internationale pour l'Exploration Scientifique de la Mer Mediterranee- CIESM, Monaco. (2008, May). The advance of thermophilic fishes in the Mediterranean Sea: overview and methodological questions. In CIESM workshop monographs (Vol. 35, pp. 39-45). CIESM, Monaco.
- 15.Balestri, E., de Battisti, D., Vallerini, F., & Lardicci, C. (2015). First evidence of root morphological and architectural variations in young Posidonia oceanica plants colonizing different substrate typologies. Estuarine, Coastal and Shelf Science, 154, 205-213. https://doi.org/10.1016/j.ecss.2015.01.002
- 16.Ballesteros, E. Mediterranean coralligenous assemblages: A synthesis of present knowledge. Oceanogr. Mar. Biol. Annu. Rev. 2006, 44, 123– 195.
- 17.Barale, V., 2008. The European marginal and enclosed seas: an overview. In: Barale, V., Gade, M. (Eds.), Remote sensing of the European seas. Springer, Dordrecht, pp. 3–22. https://doi.org/10.1007/978-1-4020-6772-3_1.

- 18.Bar-On YM, Phillips R, Milo R (2018) The biomass distribution on Earth PNAS 115:6506–6511.
- 19.Baselga, A. (2010), Partitioning the turnover and nestedness components of beta diversity. Global Ecology and Biogeography, 19: 134-143. https://doi.org/10.1111/j.1466-8238.2009.00490.x
- 20.Bell, J. D. & Harmelin-Vivien, M. I. (1982). Fish fauna of French Mediterranean Posidonia oceanica seagrass meadows. 1. Community structure. Tethys 10, 337–347.
- 21.Bell, P.R.F., Elmetri, I. & Lapointe, B.E. (2014). Evidence of Large-Scale Chronic Eutrophication in the Great Barrier Reef: Quantification of Chlorophyll a Thresholds for Sustaining Coral Reef Communities. AMBIO 43, 361–376. https://doi.org/10.1007/s13280-013-0443-1
- 22.Bellard, P. Genovesi, J.M. Jeschke Global patterns in threats to vertebrates by biological invasions P. Roy. Soc. B-Biol. Sci., 283 (1823) (2016), p. 20152454
- 23.Ben Rais Lasram F, Tomasini JA, Guilhaumon F, Romdhane MS, Do Chi T, Mouillot D (2008) Ecological correlates of dispersal success of Lessepsian fishes. Mar Ecol Prog Ser 363:273-286. https://doi.org/10.3354/meps07474
- 24.Benton, M. J. The Red Queen and the Court Jester: species diversity and the role of biotic and abiotic factors through time. Science 323, 728–732 (2009).
- 25.Berkström, C., Lindborg, R., Thyresson, M., & Gullström, M. (2013). Assessing connectivity in a tropical embayment: Fish migrations and seascape ecology. Biological Conservation, 166, 43-53. https://doi.org/10.1016/j.biocon.2013.06.013
- 26.Bevilacqua, S., Airoldi, L., Ballesteros, E., Benedetti-Cecchi, L., Boero, F., Bulleri, F., Cebrian, E., Cerrano, C., Claudet, J., Colloca, F., Coppari, M., Di Franco, A., Fraschetti, S., Garrabou, J., Guarnieri, G., Guerranti, C., Guidetti, P., Halpern, B. S., Katsanevakis, S., . . . Terlizzi, A. (2021). Mediterranean rocky reefs in the Anthropocene: Present status and future concerns. Advances in Marine Biology, 89, 1-51. https://doi.org/10.1016/bs.amb.2021.08.001
- 27.Blainey, P., Krzywinski, M. & Altman, N. Replication. Nat Methods 11, 879–880 (2014). https://doi.org/10.1038/nmeth.3091

- 28.Bleeker, A., Hicks, W. K., Dentener, F., Galloway, J., & Erisman, J. W. (2011). N deposition as a threat to the World's protected areas under the Convention on Biological Diversity. Environmental pollution, 159(10), 2280-2288. https://doi.org/10.1016/j.envpol.2010.10.036
- 29.Bolgan, M., Di Iorio, L., Dailianis, T., Catalan, I.A., Lejeune,
 P., Picciulin, M. et al. (2022). Fish acoustic community structure in Neptune seagrass meadows across the Mediterranean basin. Aquatic Conservation: Marine and Freshwater
 Ecosystems, 32(2), 329–347. https://doi.org/10.1002/aqc.3764
- 30.Bortone, S.A., & Mille, K.J. (1999). Data needs for assessing marine reserves with an emphasis on estimating fish size in situ. Il Naturalista Siciliano, Vol. XXIII, Suppl., 13±31.
- 31.Boudouresque, C.F. Marine biodiversity in the Mediterranean: Status of species, populations and communities. Sci. Rep. Port-Cros Natl. Park c, 20, 97–146.
- 32.Buñuel, X., Alcoverro, T., Pagès, J.F. et al. The dominant seagrass herbivore Sarpa salpa shifts its shoaling and feeding strategies as they grow. Sci Rep 10, 10622 (2020). https://doi.org/10.1038/s41598-020-67498-1
- 33.Busalacchi, Barbara & Rinelli, Paola & De Domenico, Francesca & Profeta, Adriana & Perdichizzi, F. & Bottari, Teresa. (2010). Analysis of demersal fish assemblages off the Southern Tyrrhenian Sea (Central Mediterranean). Hydrobiologia. 654. 111-124. 10.1007/s10750-010-0374-9.
- 34.Caldwell ZR, Zgliczynski BJ, Williams GJ, Sandin SA (2016) Reef Fish Survey Techniques: Assessing the Potential for Standardizing Methodologies. PLoS ONE 11(4): e0153066. doi: 10.1371/journal.pone.0153066
- 35.Cardinale BJ, et al. (2011) The functional role of producer diversity in ecosystems. Am J Bot 98(3):572–592.
- 36.Cardinale, B.J., Gross, K., Fritschie, K., Flombaum, P., Fox, J.W., Rixen, C., van Ruijven, J., Reich, P.B., Scherer-Lorenzen, M., Wilsey, B.J., 2013. Biodiversity simultaneously enhances the production and stability of community biomass, but the effects are independent. Ecology 94, 1697–1707. https://doi.org/10.1890/12-1334.1.
- 37.Cardinale, M., F. Colloca & G.D. Ardizzone. (1997) Feeding ecology of Mediterranean razorfish Xyrichthys novacula in the Tyrrhenian Sea (Central Mediterranean Sea). J. Appl. Ichthyol. 13: 105–111

- 38.Carlucci R, Bandelj V, Ricci P, Capezzuto F and others (2018) Exploring spatio-temporal changes in the demersal and benthopelagic assemblages of the north-western Ionian Sea (central Mediterranean Sea). Mar Ecol Prog Ser 598:1-19. https://doi.org/10.3354/meps12613.
- 39.Carminatto, A. A., Rotundo, M. M., Butturi-Gomes, D., Barrella, W., & Petrere Junior, M. (2020). Effects of habitat complexity and temporal variation in rocky reef fish communities in the Santos estuary (SP), Brazil. Ecological Indicators, 108, 105728. https://doi.org/10.1016/j.ecolind.2019.105728
- 40.Carvalho N, Afonso P, Santos RS (2003) The haremic mating system and mate choice in the Wide-eyed flounder, Bothus podas. Environ Biol Fishes 66:249–258
- 41.Castriota, L., Scarabello, M.P., Finoia, M.G. et al. (2005) Food and feeding habits of pearly razorfish, Xyrichtys novacula (Linnaeus, 1758), in the southern Tyrrhenian Sea: variation by sex and size. Environ Biol Fish 72, 123–133. https://doi.org/10.1007/s10641-004-6576-0
- 42.Cheal, A. J., Emslie, M. J., Currey-Randall, L. M., & Heupel, M. R. (2021). Comparability and complementarity of reef fish measures from underwater visual census (UVC) and baited remote underwater video stations (BRUVS). Journal of Environmental Management, 289, 112375.
- 43.Chefaoui, RM, Duarte, CM, Serrão, EA (2018). Dramatic loss of seagrass habitat under projected climate change in the Mediterranean Sea. Glob Change

```
Biol.; 24: 4919–4928. https://doi.org/10.1111/gcb.14401
```

- 44.Clarkson, E. and Beseres Pollack, J. (2021), Characterizing Fish– Habitat Associations through Paired Fisheries-Independent and In Situ Habitat Assessments. North Am. J. Fish. Manage., 41: 49-63. https://doi.org/10.1002/nafm.10530
- 45.Claudet, J., Pelletier, D., Jouvenel, J., Bachet, F., & Galzin, R. (2006). Assessing the effects of marine protected area (MPA) on a reef fish assemblage in a northwestern Mediterranean marine reserve: Identifying community-based indicators. Biological Conservation, 130(3), 349-369. https://doi.org/10.1016/j.biocon.2005.12.030
- 46.Claudet, J., Loiseau, C., Sostres, M., & Zupan, M. (2020).Underprotected marine protected areas in a global biodiversity hotspot. One Earth, 2(4), 380-384.

- 47.CMEMS, http://marine.copernicus.eu
- 48.Coll M, Piroddi C, Steenbeek J, Kaschner K, Ben Rais Lasram F, et al. (2010) The Biodiversity of the Mediterranean Sea: Estimates, Patterns, and Threats. PLoS ONE 5(8): e11842. doi: 10.1371/journal.pone.0011842.
- 49. Colton M, Swearer S (2010) A comparison of two survey methods: differences between underwater visual census and baited remote underwater video. Mar Ecol Prog Ser 400:19–36.
- 50.Da Silva, PG, Hernández, MIM, Heino, J. (2018) Disentangling the correlates of species and site contributions to beta diversity in dung beetle assemblages. Divers

Distrib; 24: 1674–1686. https://doi.org/10.1111/ddi.12785

- 51.Danovaro R., Gambi C., Dell'Anno A., Corinaldesi C., Fraschetti S., Vanreusel A., Vincx M., Gooday A.J., Exponential Decline of Deep-Sea Ecosystem Functioning Linked to BenthicBiodiversity Loss, Current Biology, Volume 18, Issue 1, 2008, Pages 1-8, ISSN 0960-9822, https://doi.org/10.1016/j.cub.2007.11.056.
- 52.De Girolamo, M., & Mazzoldi, C. (2001). The application of visual census on Mediterranean rocky habitats. Marine Environmental Research, 51(1), 1-16. https://doi.org/10.1016/S0141-1136(00)00028-3
- 53.Dennis, B., Ponciano, J.M. and Taper, M.L. (2010), Replicated sampling increases efficiency in monitoring biological populations. Ecology, 91: 610-620. https://doi.org/10.1890/08-1095.1
- 54.Deudero, S., Morey, G., Frau, A., Moranta, J. & Moreno, I. (2008). Temporal trends of littoral fishes at deep Posidonia oceanica seagrass meadows in a temperate coastal zone. Journal of Marine Systems 70, 182–195.
- 55.Di Franco A, Bussotti S, Navone A, Panzalis P, Guidetti P (2009) Evaluating effects of total and partial restrictions to fishing on Mediterranean rocky-reef fish assemblages. Mar Ecol Prog Ser 387:275-285. https://doi.org/10.3354/meps08051
- 56.Di Franco, E., Di Franco, A., Calò, A., Di Lorenzo, M., Mangialajo, L., Bussotti, S., Bianchi, C. N., & Guidetti, P. (2021). Inconsistent relationships among protection, benthic assemblage, habitat complexity and fish biomass in Mediterranean temperate rocky reefs. Ecological Indicators, 128, 107850. https://doi.org/10.1016/j.ecolind.2021.107850
- 57.Di Lorenzo, M, Guidetti, P, Di Franco, A, Calò, A, Claudet, J. Assessing spillover from marine protected areas and its drivers: A

meta-analytical approach. Fish

Fish. 2020; 21: 906-915. https://doi.org/10.1111/faf.12469

- 58.Díaz S., Pascual U., Stenseke M., Martín-López B., Watson R.T., Molnár Z., Hill R., Chan K.M.A., Baste I.A., Brauman K.A., Polasky S., Church A., Lonsdale M., Larigauderie A., Leadley P.W., Van Oudenhoven A.P.E., Van Der Plaat F., Schröter M., Lavorel S., Aumeeruddy-Thomas Y., Bukvareva E., Davies K., Demissew S., Erpul G., Failler P., Guerra C.A., Hewitt C.L., Keune H., Lindley S., Shirayama Y. 2018. Assessing nature s contributions to people: Recognizing culture, and diverse sources of knowledge, can improve assessments. SCIENCE. 359. (6373) 270-272. DOI (10.1126/science.aap8826)
- 59.Diaz-Gil, C., Grau, A., Grau, A. M., Palmer, M., Cabrera Castro, R., Jordà, G., & Catalán, I. (2019). Changes in the juvenile fish assemblage of a Mediterranean shallow Posidonia oceanica seagrass nursery area after half a century. Dickens, L.C., Goatley, C.H.R., Tanner, J.K., Bellwood, D.R., 2011. Quantifying relative diver effects in underwater visual censuses. PLoS ONE 6, e18965. https://doi.org/ 10.1371/journal.pone.0018965.
- 60.Dickens, L. C., R. Goatley, C. H., Tanner, J. K., & Bellwood, D. R. (2011). Quantifying Relative Diver Effects in Underwater Visual Censuses. PLOS ONE, 6(4), e18965. https://doi.org/10.1371/journal.pone.0018965
- 61.Duarte, C. M., Agustí, S., Kennedy, H., & Vaqué, D. (1999). The Mediterranean climate as a template for Mediterranean marine ecosystems: the example of the northeast Spanish littoral. Progress in Oceanography, 44(1-3), 245-270. https://doi.org/10.1016/S0079-6611(99)00028-2
- 62.Duffy J. E., Lefcheck J. S., Stuart-Smith R. D., Navarrete S. A., Edgar G. J., Biodiversity enhances reef fish biomass and resistance to climate change. Proc. Natl. Acad. Sci. U.S.A. 113, 6230–6235 (2016).
- 63.Edgar, G. J., Bates, A. E., Bird, T. J., Jones, A. H., Kininmonth, S., Stuart-Smith, R. D., & Webb, T. J. (2016). New approaches to marine conservation through the scaling up of ecological data. Annual Review of Marine Science, 8(1), 435–461. https://doi.org/10.1146/annurev-marine-122414-033921

- 64.Edgar, G.J., Barrett, N.S., Morton, A.J. (2004). Biases associated with the use of underwater visual census techniques to quantify the density and size-structure of fish populations.
- 65.Emslie, M.J., Cheal, A.J., MacNeil, M.A., Miller, I.R., Sweatman, H.P.A., 2018. Reef fish communities are spooked by scuba surveys and may take hours to recover. PeerJ 6, e4886. https://doi.org/10.7717/peerj.4886.
- 66.Esmaeili S., Y., N. Corte, G., Checon, H. H., G. Bilatto, C., Lefcheck, J. S., Zacagnini Amaral, A. C., & Turra, A. (2022). Revealing the drivers of taxonomic and functional diversity of nearshore fish assemblages: Implications for conservation priorities. Diversity and Distributions, 28, 1597–1609. https://doi.org/10.1111/ddi.13453
- 67.European Commission. (2020). Communication from the commission to the European parliament, the council, the European economic and social committee and the committee of the regions: EU Biodiversity Strategy for 2030. Bringing nature back into our lives.
- 68.Fasulo, S., Marino, S., Mauceri, A., Maisano, M., Giannetto, A., D'Agata, A., Parrino, V., Minutoli, R., & De Domenico, E. (2010). A multibiomarker approach in Coris julis living in a natural environment. Ecotoxicology and Environmental Safety, 73(7), 1565-1573. https://doi.org/10.1016/j.ecoenv.2010.01.008
- 69. Fiorella, K. J., E. R. Bageant, M. Kim, V. Sean, V. Try, H. J. MacDonell, E. Baran, Y. Kura, A. C. Brooks, C. B. Barrett, and S. Thilsted. 2019. Analyzing drivers of fish biomass and biodiversity within community fish refuges in Cambodia. Ecology and Society 24(3):18. https://doi.org/10.5751/ES-11053-240318.
- 70.Ford BM, Stewart BA, Roberts JD (2017) Species pools and habitat complexity define Western Australian marine fish community composition. Mar Ecol Prog Ser 574:157-166. https://doi.org/10.3354/meps12167
- 71.Franco, A., Franzoi, P., Malavasi, S., Riccato, F., Torricelli, P., & Mainardi, D. (2006). Use of shallow water habitats by fish assemblages in a Mediterranean coastal lagoon. Estuarine, Coastal and Shelf Science, 66(1-2), 67-83. https://doi.org/10.1016/j.ecss.2005.07.020
- 72.Francour, P. (1997), Fish Assemblages of Posidonia oceanica Beds at Port-Cros (France, NW Mediterranean): Assessment of Composition and Long-Term Fluctuations by Visual Census. Marine Ecology, 18: 157-173. https://doi.org/10.1111/j.1439-0485.1997.tb00434.x

- 73.Fraija-Fernández, N, Bouquieaux, M-C, Rey, A, et al. (2020). Marine water environmental DNA metabarcoding provides a comprehensive fish diversity assessment and reveals spatial patterns in a large oceanic area. Ecol Evol. 10: 7560–7584. https://doi.org/10.1002/ece3.6482
- 74.Frasca, S., Wolf, J. C., Kinsel, M. J., Camus, A. C., & Lombardini, E. D. (2018). Osteichthyes. In Pathology of Wildlife and Zoo Animals (pp. 953-1001). Elsevier. https://doi.org/10.1016/B978-0-12-805306-5.00039-0
- 75.French, B., Wilson, S., Holmes, T., Kendrick, A., Rule, M., & Ryan, N. (2021). Comparing five methods for quantifying abundance and diversity of fish assemblages in seagrass habitat. Ecological Indicators, 124, 107415. https://doi.org/10.1016/j.ecolind.2021.107415
- 76.Froese, R. and D. Pauly. Editors. 2022.FishBase. World Wide Web electronic publication. www.fishbase.org, (06/2022)
- 77.Fruciano, C., Tigano, C., & Ferrito, V. (2011). Geographical and morphological variation within and between colour phases in Coris julis (L. 1758), a protogynous marine fish. Biological Journal of the Linnean Society, 104(1), 148-162.
- 78.Galli, M., Tepsich, P., Baini, M., Panti, C., Rosso, M., Vafeiadou, A., Pantelidou, M., Moulins, A., & Fossi, M. C. (2022). Microplastic abundance and biodiversity richness overlap: Identification of sensitive areas in the Western Ionian Sea. Marine Pollution Bulletin, 177, 113550. https://doi.org/10.1016/j.marpolbul.2022.113550
- 79.Gkafas GA, Tsigenopoulos C, Magoulas A, Panagiotaki P, Vafidis D, Mamuris Z, Exadactylos A (2013) Population subdivision of saddled seabream Oblada melanura in the Aegean Sea revealed by genetic and morphometric analyses. Aquat Biol 18:69-80. <u>https://doi.org/10.3354/ab00490</u>
- 80.Gordoa, A., & Molí, B. (1997). Age and growth of the sparids Diplodus vulgaris, D. sargus and D. annularis in adult populations and the differences in their juvenile growth patterns in the north-western Mediterranean Sea. Fisheries Research, 33(1-3), 123-129. https://doi.org/10.1016/S0165-7836(97)00074-X
- 81.Gotelli, Nicholas & Chao, Anne. (2013). Measuring and Estimating Species Richness, Species Diversity, and Biotic Similarity from Sampling Data. 10.1016/B978-0-12-384719-5.00424-X.
- 82.Gratwicke, B. and Speight, M.R. (2005), The relationship between fish species richness, abundance and habitat complexity in a range of

shallow tropical marine habitats. Journal of Fish Biology, 66: 650-667. https://doi.org/10.1111/j.0022-1112.2005.00629.x

- 83.Guidetti, P. (2000) Differences Among Fish Assemblages Associated with Nearshore Posidonia oceanica Seagrass Beds, Rocky–algal Reefs and Unvegetated Sand Habitat in the Adriatic Sea. Estuarine, Coastal and Shelf Science, Volume 50, Issue 4, 2000, Pages 515-529, ISSN 0272-7714, https://doi.org/10.1006/ecss.1999.0584.
- 84.Guidetti, P., Baiata, P., Ballesteros, E., Franco, A. D., Hereu, B., Macpherson, E., Micheli, F., Pais, A., Panzalis, P., Rosenberg, A. A., Zabala, M., & Sala, E. (2014). Large-Scale Assessment of Mediterranean Marine Protected Areas Effects on Fish Assemblages. PLOS ONE, 9(4), e91841. https://doi.org/10.1371/journal.pone.0091841
- 85.Gurevitch, J., & Padilla, D. K. (2004). Are invasive species a major cause of extinctions? Trends in Ecology & Evolution, 19(9), 470-474. https://doi.org/10.1016/j.tree.2004.07.005
- 86.Hall, A. E., & Kingsford, M. J. (2021). Habitat type and complexity drive fish assemblages in a tropical seascape. Journal of Fish Biology, 99(4), 1364–1379. https://doi.org/10.1111/jfb.14843
- 87.Harmelin-Vivien, M.L., Harmelin, J.G., Chauvet, C., Duval, C., Galzin, R., Lejeune, P., Barnab'e, G., Blanc, F., Chevalier, R., Duclerc, J., Lasserre, G., 1985. Evaluation visuelle des peuplements et populations de poissons m'ethodes et probl'emes. Soci'et'e Natl. Prot. Nat. Acclim. Fr. Paris 40, 467–539.
- 88.Harper, L. M., Lefcheck, J. S., Whippo, R., Jones, M. S., Foltz, Z., & Duffy, J. E. (2022). Blinded by the bright: How species-poor habitats contribute to regional biodiversity across a tropical seascape. Diversity and Distributions, 28, 2272–2285. https://doi.org/10.1111/ddi.13632
- 89.Heino, J., Grönroos, M. (2017). Exploring species and site contributions to beta diversity in stream insect assemblages. Oecologia 183, 151–160. https://doi.org/10.1007/s00442-016-3754-7
- 90.Henriques, S., Pais, M. P., Costa, M. J., & Cabral, H. N. (2013). Seasonal variability of rocky reef fish assemblages: Detecting functional and structural changes due to fishing effects. Journal of Sea Research, 79, 50-59. https://doi.org/10.1016/j.seares.2013.02.004

- 91.Henseler, C., Nordström, M. C., Törnroos, A., Snickars, M., Pecuchet, L., Lindegren, M., & Bonsdorff, E. (2019). Coastal habitats and their importance for the diversity of benthic communities: A species-and trait-based approach. Estuarine, Coastal and Shelf Science, 226, 1–14.
- 92.Hermoso, V., Carvalho, S. B., Giakoumi, S., Goldsborough, D., Katsanevakis, S., Leontiou, S., ... & Yates, K. L. (2022). The EU Biodiversity Strategy for 2030: Opportunities and challenges on the path towards biodiversity recovery. Environmental Science & Policy, 127, 263-271.
- 93.Hilborn, R., and Walters, C. J. Eds. 2013. Quantitative fisheries stock assessment: choice, dynamics and uncertainty. Springer Science & Business Media, Dordrecht.
- 94.Hill, R., Díaz, S., Pascual, U., Stenseke, M., Molnár, Z., & Van Velden, J. (2021). Nature's contributions to people: Weaving plural perspectives. One Earth, 4(7), 910-915. https://doi.org/10.1016/j.oneear.2021.06.009
- 95.Holm S (1979) A simple sequentially rejective multiple test procedure. Scand J Stat 6(2):65–70
- 96.Hooper, D. U., Chapin III, F. S., Ewel, J. J., Hector, A., Inchausti, P., Lavorel, S., ... & Wardle, D. A. (2005). Effects of biodiversity on ecosystem functioning: a consensus of current knowledge. Ecological monographs, 75(1), 3-35.
- 97.Hyndes, G. A., Francour, P., Guidetti, P., Heck, K. L., & Jenkins, G. (2018). The roles of seagrasses in structuring associated fish assemblages and fisheries. In Seagrasses of Australia (pp. 589-627). Springer, Cham.
- 98.Iacono, R.; Napolitano, E.; Palma, M.; Sannino, G. The Tyrrhenian Sea Circulation: A Review of Recent Work. Sustainability 2021, 13, 6371. https://doi.org/10.3390/su13116371
- 99.Jaxion-Harm, J., Saunders, J, & Speight, M. R. (2012). Distribution of fish in seagrass, mangroves and coral reefs: life-stage dependent habitat use in Honduras. Revista de Biología Tropical, 60(2), 683-698.
- 100. Jenkins, G. P., & Wheatley, M. J. (1998). The influence of habitat structure on nearshore fish assemblages in a southern Australian embayment: Comparison of shallow seagrass, reef-algal and unvegetated sand habitats, with emphasis on their importance to

recruitment. Journal of Experimental Marine Biology and Ecology, 221(2), 147-172. https://doi.org/10.1016/S0022-0981(97)00121-4

101. Jordà, G., Von Schuckmann, K., Josey, S., Caniaux, G., García-Lafuente, J., Sammartino, S., Özsoy, E., Polcher, J., Notarstefano, G., Poulain, P., Adloff, F., Salat, J., Naranjo, C., Schroeder, K., Chiggiato, J., Sannino, G., & Macías, D. (2017). The Mediterranean Sea heat and mass budgets: Estimates, uncertainties and perspectives. Progress in Oceanography, 156, 174-208.

https://doi.org/10.1016/j.pocean.2017.07.001

- 102. Kalimeris, Anastasios & Kassis, Dimitris. (2020). Sea surface circulation variability in the Ionian-Adriatic Seas. Progress in Oceanography. 189. 102454. 10.1016/j.pocean.2020.102454.
- 103. Kalogirou, S., Corsini-Foka, M., Sioulas, A., Wennhage, H. and Pihl, L. (2010), Diversity, structure and function of fish assemblages associated with Posidonia oceanica beds in an area of the eastern Mediterranean Sea and the role of non-indigenous species. Journal of Fish Biology, 77: 2338-2357. https://doi.org/10.1111/j.1095-8649.2010.02817.x
- 104. Karachle, P.K. and Stergiou, K.I. (2008), The effect of season and sex on trophic levels of marine fishes. Journal of Fish Biology, 72: 1463-1487. https://doi.org/10.1111/j.1095-8649.2008.01809.x
- 105. Kelly RP, Closek CJ, O'Donnell JL, Kralj JE, Shelton AO and Samhouri JF (2017) Genetic and Manual Survey Methods Yield Different and Complementary Views of an Ecosystem. Front. Mar. Sci. 3:283. doi: 10.3389/fmars.2016.00283
- 106. King M. Fisheries Management. Fisheries Biology, Assessment and Management. Oxford, UK: Blackwell Publishing Ltd; 2013. pp. 273–315. doi: 10.1002/9781118688038.ch6.
- Koleff, P., Gaston, K.J. and Lennon, J.J. (2003), Measuring beta diversity for presence–absence data. Journal of Animal Ecology, 72: 367-382. https://doi.org/10.1046/j.1365-2656.2003.00710.x
- 108. Kotta D, Kitsiou D. (2019). Chlorophyll in the Eastern Mediterranean Sea: Correlations with Environmental Factors and Trends. *Environments.*; 6(8):98. https://doi.org/10.3390/environments6080098
- 109. Kulbicki, M. (1998). How the acquired behaviour of commercial reef fishes may influence the results obtained from visual censuses.

Journal of Experimental Marine Biology and Ecology, 222(1-2), 11-30. https://doi.org/10.1016/S0022-0981(97)00133-0

- La Mesa, G., Molinari, A., Gambaccini, S. and Tunesi, L. (2011), Spatial pattern of coastal fish assemblages in different habitats in North-western Mediterranean. Marine Ecology, 32: 104-114. https://doi.org/10.1111/j.1439-0485.2010.00404.x
- 111. Labropoulou, M., Machias, A., Tsimenides, N. & Eleftheriou, A. (1997). Feeding habits and ontogenetic diet shift of the striped red mullet, Mullus surmuletus Linnaeus, 1758. Fisheries Research 31, 257– 267.
- 112. Lazzari, N., Martín-López, B., Sanabria-Fernandez, J. A., & Becerro, M. A. (2020). Alpha and beta diversity across coastal marine social-ecological systems: Implications for conservation. Ecological Indicators, 109, 105786. https://doi.org/10.1016/j.ecolind.2019.105786
- 113. Le Bagousse-Pinguet, Y., Soliveres, S., Gross, N., Torices, R., Berdugo, M., & Maestre, F. T. (2019). Phylogenetic, functional, and taxonomic richness have both positive and negative effects on ecosystem multifunctionality.
- 114. Legendre, P., & de Cáceres, M. (2013). Beta diversity as the variance of community data: Dissimilarity coefficients and partitioning. Ecology Letters, 16(8), 951–963. https://doi.org/10.1111/ele.12141.
- 115. Lewis, J., Farnsworth, M., Burdett, C. et al. Biotic and abiotic factors predicting the global distribution and population density of an invasive large mammal. Sci Rep 7, 44152 (2017). https://doi.org/10.1038/srep44152
- 116. Loiseau, N., & Gaertner, C. (2015). Indices for assessing coral reef fish biodiversity: the need for a change in habits. Ecology and Evolution, 5(18), 4018-4027. https://doi.org/10.1002/ece3.1619
- 117. Loreau M, Naeem S, Inchausti P, Bengtsson J, Grime JP, Hector A, Hooper DU, Huston MA, Raffaelli D, Schmid B, Tilman D, Wardle DA. Biodiversity and ecosystem functioning: current knowledge and future challenges. Science. 2001 Oct 26;294(5543):804-8. doi: 10.1126/science.1064088. PMID: 11679658.
- 118. Lowry, M., Folpp, H., Gregson, M., Suthers, I., 2012. Comparison of baited remote underwater video (BRUV) and underwater visual census (UVC) for assessment of artificial reefs in estuaries. J. Exp. Mar. Biol. Ecol. 416–417, 243–253.
- 119. Madi Moussa R., Bertucci F., Jorissen H., Gache C., Pita Waqalevu V., Parravicini V., Lecchini D., Galzin R., Importance of
intertidal seagrass beds as nursery area for coral reef fish juveniles (Mayotte, Indian Ocean), Regional Studies in Marine Science, Volume 33, 2020, 100965, ISSN 2352-4855,

https://doi.org/10.1016/j.rsma.2019.100965.

- 120. Magurran, A.E. (2004) Measuring Biological Diversity. Blackwell Publishing, Oxford, 256 p.
- 121. Mallet, D., Pelletier, D., 2014. Underwater video techniques for observing coastal marine biodiversity: a review of sixty years of publications (1952–2012). Fish. Res. 154, 44–62.
- 122. Mallet, D.; Olivry, M.; Ighiouer, S.; Kulbicki, M.; Wantiez, L. Nondestructive Monitoring of Soft Bottom Fish and Habitat Using a Standardized, Remote and Unbaited 360° Video Sampling Method. Fishes 2021, 6, 50. https://doi.org/10.3390/ fishes6040050
- 123. Mammola, S., Riccardi, N., Prié, V., Correia, R., Cardoso, P., Lopes-Lima, M., & Sousa, R. (2020). Towards a taxonomically unbiased European Union biodiversity strategy for 2030. Proceedings of the Royal Society B, 287(1940), 20202166.
- Manel, S., Guerin, PE., Mouillot, D. et al. Global determinants of freshwater and marine fish genetic diversity. Nat Commun 11, 692 (2020). https://doi.org/10.1038/s41467-020-14409-7
- Marbà, N., Díaz-Almela, E., & Duarte, C.
 M. (2014). Mediterranean seagrass (Posidonia oceanica) loss between 1842 and 2009. Biological Conservation, 176, 183-190. https://doi.org/10.1016/j.biocon.2014.05.024
- 126. Martin S., Aspden R.J., and. Paterson D.M. (eds), Marine Biodiversity and Ecosystem Functioning: Frameworks, methodologies, and integration (Oxford, 2012; online edn, Oxford Academic, 17 Dec. 2013), https://doi.org/10.1093/acprof:oso/9780199642250.001.0001
- 127. Matic-Skoko, S., Ferri, J., Kraljevic, M., & Dulcic, J. (2007). Growth of juvenile striped seabream, Lithognathus mormyrus (Teleostei: Sparidae), in the Adriatic Sea. Journal of Applied Ichthyology, 23(3), 286.
- 128. McCauley DJ, Pinsky ML, Palumbi SR, Estes JA, Joyce FH, Warner RR. Marine defaunation: animal loss in the global ocean. Science. 2015 Jan 16;347(6219):1255641. doi: 10.1126/science.1255641. PMID: 25593191.
- 129. McClure, E.C., Sievers, K.T., Abesamis, R.A., Hoey, A.S., Alcala, A.C., Russ, G.R., 2020. Higher fish biomass inside than outside marine protected areas despite typhoon impacts in a complex reefscape.

Biol. Conserv. 241, 108354. https://doi.org/ 10.1016/j.biocon.2019.108354.

Meier, E.S., Kienast, F., Pearman, P.B., Svenning, J.-C., Thuiller, W., Araújo, M.B., Guisan, A. and Zimmermann, N.E. (2010), Biotic and abiotic variables show little redundancy in explaining tree species distributions. Ecography, 33: 10381048, https://llii.org/10.1111/j.1600.0587.2010.06220

1048. https://doi.org/10.1111/j.1600-0587.2010.06229.x

- Micheli F, Halpern BS, Walbridge S, Ciriaco S, Ferretti F, et al. (2013) Cumulative Human Impacts on Mediterranean and Black Sea Marine Ecosystems: Assessing Current Pressures and Opportunities. PLoS ONE 8(12): e79889. doi:10.1371/journal.pone.0079889
- 132. Midgley GF (2012) Biodiversity and ecosystem function. Science 335:174–175.
- 133. Moranta, J., Palmer, M., Morey, G., Ruiz, A. & Morales-Nin, B. (2006). Multi-scale spatial variability in fish assemblages associated with Posidonia oceanica meadows in the Western Mediterranean Sea. Estuarine, Coastal and Shelf Science 68, 579–592.
- Mouillot, D., Albouy, C., Guilhaumon, F., Ben Rais Lasram, F., Coll, M., Devictor, V., Meynard, C. N., Pauly, D., Tomasini, J. A., Troussellier, M., Velez, L., Watson, R., Douzery, E. J., & Mouquet, N. (2011). Protected and Threatened Components of Fish Biodiversity in the Mediterranean Sea. Current Biology, 21(12), 1044-1050. https://doi.org/10.1016/j.cub.2011.05.005
- 135. Nicolaidou, A., Alongi, G., Aydogan, O., Catra, M., Cavas, L., Cevik, C., ... & Filiz, H. (2012). New mediterranean biodiversity records (June 2012).
- 136. Pearson, R. G. & Dawson, T. P. Predicting the impacts of climate change on the distribution of species: are bioclimate envelope models useful? Global Ecology and Biogeography 12, 361–371 (2003).
- 137. Pennino M.G., Conesa D., López-Quílez A., Muñoz F, Fernández A., Bellido J.M., (2016) Fishery-dependent and -independent data lead to consistent estimations of essential habitats, ICES Journal of Marine Science, Volume 73, Issue 9, Pages 2302–2310, https://doi.org/10.1093/icesjms/fsw062
- Pergent-Martini, C., Boudouresque, C.-F., Pasqualini, V. and Pergent, G. (2006), Impact of fish farming facilities on Posidonia oceanica meadows: a review. Marine Ecology, 27: 310-319. https://doi.org/10.1111/j.1439-0485.2006.00122.x

- 139. Perry, D., Staveley, T. A., & Gullström, M. (2018). Habitat Connectivity of Fish in Temperate Shallow-Water Seascapes. Frontiers in Marine Science, 4. https://doi.org/10.3389/fmars.2017.00440
- 140. Piazzi, L., Cecchi, E., Serena, F.(2012). Spatial and temporal patterns of diversity in mediterranean rocky reef fish assemblages. Vie et Milieu. 62. 129-136.
- 141. Pinnegar, J.K. (2018), Why the damselfish Chromis chromis is a key species in the Mediterranean rocky littoral a quantitative perspective. J Fish Biol, 92: 851-872. https://doi.org/10.1111/jfb.13551
- 142. Plant, T. M., & Zeleznik, A. J. (Eds.). (2014). Knobil and Neill's physiology of reproduction. Academic Press.
- Polanco Fernández, A, Marques, V, Fopp, F, et al.
 (2020) Comparing environmental DNA metabarcoding and underwater visual census to monitor tropical reef fishes. Environmental DNA. 2021; 3: 142–156. https://doi.org/10.1002/edn3.140
- 144. Poulos, S.E. (2020). The mediterranean and black sea marine system: An overview of its physico-geographic and oceanographic characteristics. Earth Sci. Rev. 200, 103004.
- 145. Raitsos D. E., Beaugrand G., Georgopoulos D., Zenetos
 A., Pancucci-Papadopoulou A. M., Theocharis A., Papathanassiou E., (2010), Global climate change amplifies the entry of tropical species into the eastern Mediterranean Sea, Limnology and Oceanography, 55, doi: 10.4319/lo.2010.55.4.1478.
- 146. Ramirez-Llodra E, Tyler PA, Baker MC, Bergstad OA, Clark MR, et al. (2011) Man and the Last Great Wilderness: Human Impact on the Deep Sea. PLOS ONE 6(8): e22588. https://doi.org/10.1371/journal.pone.0022588
- 147. Rattray A, Andrello M, Asnaghi V, Bevilacqua S and others (2016) Geographic distance, water circulation and environmental conditions shape the biodiversity of Mediterranean rocky coasts. Mar Ecol Prog Ser 553:1-11. https://doi.org/10.3354/meps11783
- 148. Reiss, C. S., Panteleev, G., Taggart, C. T., Sheng, J., & DeYoung, B. (2000). Observations on larval fish transport and retention on the Scotian Shelf in relation to geostrophic circulation. Fisheries Oceanography, 9(3), 195-213.
- 149. Rice J., Environmental health indicators, Ocean & Coastal Management, Volume 46, Issues 3–4, 2003, Pages 235-259, ISSN 0964-5691

- 150. Rodrigues, L. d. S., Pennino, M. G., Conesa, D., Kikuchi, E., Kinas, P. G., Barbosa, F. G., & Cardoso, L. G. (2022). Modelling the distribution of marine fishery resources: Where are we? Fish and Fisheries, 00, 1–17. https://doi.org/10.1111/faf.12716
- 151. Sakalli, A. (2017). Sea surface temperature in the Mediterranean Sea under climate change: a linear model simulation of the sea surface temperature up to 2100. Appl. Ecol. Environ. Res. 15, 707–716. doi: 10.15666/aeer/1501_707716
- 152. Sala, E. and E. Ballesteros (1997). Partitioning of space and food resources by three fish genus Diplodus (Sparidae) in a Mediterranean rocky infralittoral ecosystem. Mar. Ecol. Prog. Ser. 152:273-283.
- 153. Sala, E., Ballesteros, E., Dendrinos, P., Franco, A. D., Ferretti, F., Foley, D., Fraschetti, S., Friedlander, A., Garrabou, J., Güçlüsoy, H., Guidetti, P., Halpern, B. S., Hereu, B., Karamanlidis, A. A., Kizilkaya, Z., Macpherson, E., Mangialajo, L., Mariani, S., Micheli, F., . . . Zabala, M. (2012). The Structure of Mediterranean Rocky Reef Ecosystems across Environmental and Human Gradients, and Conservation Implications. PLOS ONE, 7(2), e32742. https://doi.org/10.1371/journal.pone.0032742
- 154. Sale PF (1980) The ecology of fishes on coral reefs.Oceanography and Marine Biology an Annual Review 18: 367-442
- 155. Schintu P, Passariello M, Belluscio A, Ardizzone GD (1994) Growth and diet of Bothus podas (Pisces: Bothidea) in the Central Méditerranean Sea. Sci Mar 58:359–361
- 156. Schramm, K. D., Marnane, M. J., Elsdon, T. S., Jones, C. M., Saunders, B. J., Newman, S. J., & Harvey, E. S. (2021). Fish associations with shallow water subsea pipelines compared to surrounding reef and soft sediment habitats. Scientific Reports, 11(1), 1–15. https://doi.org/10.1038/s4159 8-021- 85396
- 157. Seitz R. D., Wennhage H., Bergström U., Lipcius R.N., Ysebaert T., Ecological value of coastal habitats for commercially and ecologically important species, ICES Journal of Marine Science, Volume 71, Issue 3, March/April 2014, Pages 648–665, https://doi.org/10.1093/icesjms/fst152.
- 158. Shaltout, M., & Omstedt, A. (2014). Recent sea surface temperature trends and future scenarios for the Mediterranean Sea. Oceanologia, 56(3), 411-443. https://doi.org/10.5697/oc.56-3.411
- 159. Shannon, C.E., Weaver W.(1949). The mathematical theory of communication, University of Illinois Press, Illinois.

- 160. Shepherd, B., Pinheiro, H. T., Phelps, A. Y., Pérez-Matus, A., & Rocha, L. A. (2021). Pseudanthiashangapiko, a new anthiadine serranid (Teleostei, Serranidae, Anthiadinae) from Rapa Nui (Easter Island). ZooKeys, 1054, 1-13. https://doi.org/10.3897/zookeys.1054.64508
- 161. Sieber, A., & Pyle, R. (2010). A review of the use of closedcircuit rebreathers for scientific diving. Underwater Technology, 29(2), 73-78.
- 162. Simpson E.H (1949). Measurement of diversity, Nature, 163, p. 688.
- 163. Spanò, N., De Domenico, E.. "Biodiversity in Central Mediterranean Sea". Mediterranean Identities - Environment, Society, Culture, edited by Borna Fuerst-Bjelis, IntechOpen, 2017. 10.5772/intechopen.68942.
- 164. Sperone, E., Parise, G., Leone, A., Milazzo, C., Circosta, V., Santoro, G., ... & Tripepi, S. (2012). Spatiotemporal patterns of distribution of large predatory sharks in Calabria (central Mediterranean, southern Italy). Acta Adriatica, 53(1), 13-24.
- 165. Staveley, T.A.B., Perry, D., Lindborg, R. and Gullström, M. (2017), Seascape structure and complexity influence temperate seagrass fish assemblage composition. Ecography, 40: 936-946. https://doi.org/10.1111/ecog.02745
- 166. Stuart-Smith, R., Bates, A., Lefcheck, J. et al. Integrating abundance and functional traits reveals new global hotspots of fish diversity. Nature 501, 539–542 (2013). https://doi.org/10.1038/nature12529
- 167. Telesca, L., Belluscio, A., Criscoli, A., Ardizzone, G., Apostolaki, E.T., Fraschetti, S., Gristina, M., Knittweis, L., Martin, C.S., Pergent, G. and Alagna, A., 2015. Seagrass meadows (Posidonia oceanica) distribution and trajectories of change. Sci. Rep. 5, 12505.
- 168. Thiriet, P.D., Di Franco, A., Chemin'ee, A., Guidetti, P., Bianchimani, O., Basthard- Bogain, S., Cottalorda, J.-M., Arceo, H., Moranta, J., Lejeune, P., Francour, P., Mangialajo, L., 2016. Abundance and diversity of crypto- and necto-benthic coastal fish are higher in marine forests than in structurally less complex macroalgal assemblages. PLOS ONE 11, e0164121. https://doi.org/10.1371/journal. pone.0164121.
- 169. Thresher RE, Gunn JS (1986) Comparative analysis of visual census techniques for highly mobile, reef-associated piscivores (Carangidae). Environmental Biology of Fishes 17: 93-116

- 170. Thuiller, W., B. Lafourcade, R. Engler, and M. B. Araujo.(2009). "BIOMOD a Platform for Ensemble Forecasting of Species Distributions." Ecography 32: 369–73
- 171. Tiedemann, M., Fock, H. O., Döring, J., Badji, L. B., & Möllmann, C. (2018). Water masses and oceanic eddy regulation of larval fish assemblages along the Cape Verde Frontal Zone. Journal of Marine Systems, 183, 42-55.

https://doi.org/10.1016/j.jmarsys.2018.03.004

- 172. Tortonese E 1970. Fauna d'italia osteichthyes pesci ossei. Bologna: Ed. Calderini.
- 173. Trabelsi, A., & Rassoulzadegan, F. (2011). Effect of bacterial community dynamics on DOC seasonal changes in the north-western Mediterranean Sea. Journal of plankton research, 33(8), 1249-1262.
- Tserpes, G., Fiorentino, F., Levi, D., Cau, A., Murenu,
 M., Zamboni, A. & Papaconstantinou, C. (2002). Distribution of Mullus barbatus and M. surmuletus (Osteichthyes: Perciformes) in the Mediterranean continental shelf: implications for management. Scientia Marina 66, 29–54.
- 175. Uiblein, F. (2007). Goatfishes (Mullidae) as indicators in tropical and temperate coastal habitat monitoring and management. Marine Biology Research 3, 275–288.
- 176. UN General Assembly, Transforming our world : the 2030 Agenda for Sustainable Development, 21 October 2015, A/RES/70/1, available at: https://www.refworld.org/docid/57b6e3e44.html
- 177. Van der Putten, W. H., Macel, M. & Visser, M. E. Predicting species distribution and abundance responses to climate change: why it is essential to include biotic interactions across trophic levels. Philosophical Transactions of the Royal Society B: Biological Sciences 365, 2025–2034 (2010).
- 178. Vega Fernández T, D'Anna G, Badalamenti F, Pérez-Ruzafa A (2008) Habitat connectivity as a factor affecting fish assemblages in temperate reefs. Aquat Biol 1:239-248. https://doi.org/10.3354/ab00027
- 179. Verberk, W. C. E. P. (2011). Explaining general patterns in species abundance and distributions. Nature education knowledge, 3(10), 38.
- 180. Waycott M, Duarte CM, Carruthers TJ, Orth RJ, Dennison WC, Olyarnik S, Calladine A, Fourqurean JW, Heck KL, Hughes AR+1

more. 2009. Accelerating loss of seagrasses across the globe threatens coastal ecosystems. Proceedings of the National Academy of Sciences of the United States of America 106:12377-12381

- 181. Whittaker, R. H. (1960) Vegetation of the Siskiyou Mountains, Oregon and California. Ecological Monographs, 30, 279–338
- 182. Wiens, J. J. The niche, biogeography and species interactions. Philosophical Transactions of the Royal Society of London B: Biological Sciences 366, 2336–2350 (2011).
- 183. Williams, R. G., & Follows, M. J. (2003). Physical transport of nutrients and the maintenance of biological production. In Ocean biogeochemistry (pp. 19-51). Springer, Berlin, Heidelberg.
- 184. Worm, B. et al. Impacts of biodiversity loss on ocean ecosystem services. Science 314, 787–790 (2006).
- 185. Yancey PH, Gerringer ME, Drazen JC, Rowden AA, Jamieson A. Marine fish may be biochemically constrained from inhabiting the deepest ocean depths. Proc Natl Acad Sci U S A. 2014 Mar 25;111(12):4461-5. doi: 10.1073/pnas.1322003111.
- 186. Zenetos A, Ertan M, Crocetta F, Golani D, Rosso A, Servello G, Shenkar N, Turon X, Verlaque M (2017) Uncertainties and validation of alien species catalogues : The Mediterranean as an example. Estuarine, Coastal and Shelf Science 191: 171–187, https://doi.org/10.1016/j.ecss. 2017.03.031
- 187. Zhao, W.; Zhang, X.; Jia, G.; Shen, Y.; Zhu, M. (2021). "The Silurian-Devonian boundary in East Yunnan (South China) and the minimum constraint for the lungfish-tetrapod split". Science China Earth Sciences. 64 (10): 1784–1797.
- 188. Zuur, A.F., Ieno, E.N. and Elphick, C.S. (2010), A protocol for data exploration to avoid common statistical problems. Methods in Ecology and Evolution, 1: 3-14. https://doi.org/10.1111/j.2041-210X.2009.00001.x.